



## A state-of-the-art review on dual purpose seaweeds utilization for wastewater treatment and crude bio-oil production

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### ABSTRACT

Due to the negative environmental impacts of fossil fuels and the increasing global energy demands, biofuels are receiving increasing attention as the best short-term substitute for petroleum. Recently, thermochemical conversion of seaweeds is in industrial focus to obtain high-value products with more potential applications than the conventional raw material. Beside biofuel production and due to their autotrophic growth, seaweeds are receiving a great attention in the field of bioremediation. Thus, pyrolysis of seaweeds is a promising approach for renewable bio-oil production with positive environmental impacts. However, a pretreatment drying step is required to improve the conversion process of the biomass. Application of electro-osmotic dewatering as well as on-site mechanical dewatering methods prior to the drying process were reported as useful techniques to reduce the energy requirements. On the other hand, the bio-oil produced from pyrolysis of seaweeds usually has high contents of oxygen-, nitrogen- and sulphur-containing compounds, which should be as minimum as possible to enhance the bio-oil stability and reduce NO<sub>x</sub> and SO<sub>x</sub> emissions. The present review introduces a suggested route combining a number of technologies that create an economically-feasible process for conversion of seaweeds to high-grade crude bio-oil through pyrolysis. In addition, the paper sheds light on the environmental impacts and economic feasibility of the crude bio-oil production from seaweeds. The current status and challenges related to pyrolysis, as well as future perspectives for enhanced conversion and upgraded bio-oil production, are discussed.

### 1. Introduction

The growing industrialization and human population during the recent decades resulted in potential increase in the global energy demands. The current petroleum consumption rate is estimated by about 10<sup>5</sup> times faster than the nature can create [1]. If this energy consumption rate continues, it is predicted that the world will face an energy crisis due to exhaustion of the worldwide fossil oil reserves in shorter than 3 decades [2]. In addition, dependence on fossil oil as a main energy source contributes to excessive CO<sub>2</sub> emission [3], with about 20% of the worldwide CO<sub>2</sub> emissions from transportation sector only [4]. Taking this sector as an example, the total new-vehicle annual sales in 2013 were 84 million, which is expected to increase to 127 million by 2035, bringing the total global vehicle number to 2 billion

[5]. CO<sub>2</sub> is the main contributor for the global warming, which is the 'talk of the town' all over the world as a great threat to mankind and the planet. In addition to the significant changes in weather patterns, global warming results in potential increase in the sea level and consequently flooding of lowlands and islands. Therefore, continuous reliance on petroleum is now widely documented to be unsustainable. Replacement of fossil-based energy with green renewable resources has received much attention globally from research sectors as well as governments and industry. Amongst, different biofuels have been discussed as a biomass-derived fuels which are renewable, sustainable, and eco-friendly alternatives to petroleum. Together with other renewable energy sources, biofuels have the potential to completely replace the current conventional energy sources, reinforcing energy security to reduce the emissions of the greenhouse gases (GHGs).

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Photoautotrophic microorganisms are considered as a promising candidate for unceasing energy appetite. Recently, more interest has been devoted to the third-generation biofuel feedstocks, representing algae, because first-generation edible feedstocks are under serious controversy due to the competition with human food [6]. In addition, conversion of second-generation biofuel feedstocks, such as lignocellulosic wastes, is limited due to the elevated cost of lignin degradation [7]. Therefore, algae-based biofuels progressed wildly in the last decades as one of the most important renewable energy research directions for countering these issues. Algae, including micro- and macroalgae/seaweeds, are photoautotrophic organisms with high photosynthetic efficiency of 6–8%, which reported to be much higher than that of terrestrial plants (1.8–2.2%) [8]. In addition, algae are very efficient in utilizing the nutrients from wastewater [9] or seawater [10], and they don't need arable land to grow [11]. Algal biomass can be converted into different forms of bioenergy such as crude bio-oil, biogas, bioethanol and biodiesel through different routes including thermochemical conversion, anaerobic digestion, fermentation and lipid transesterification, respectively (Table 1).

Seaweeds have been explored as a biofuel feedstock, which needs further research to fully explore their potential for crude bio-oil production. Due to the low lipid content of seaweeds in comparison to microalgae, they are widely discussed as a high potential feedstock for crude bio-oil production through different methods of thermochemical conversion. Comparing to terrestrial plants, the annual biomass productivity of seaweeds is much higher, and at the same time is much easier to maintain and harvest compared to microalgae [19]. According to a Life Cycle Assessment (LCA) carried by Aitken et al. [34], seaweeds can generate a net energy of 11.0 GJ ton<sup>-1</sup> of dry weight (dw) compared to 9.5 GJ ton<sup>-1</sup> dw of microalgae. However, applications of seaweeds for crude bio-oil production are at infancy stage and need economically-efficient technological solutions [35,36]. So far, there is limited review papers to evaluate seaweeds conversion into crude bio-oil, especially, via pyrolysis of seaweeds cultivated in wastewater effluents for dual propose of energy production and phycoremediation. Therefore, the objective of this review is to discuss and provide up-to-date knowledge on seaweeds utilization for dual purpose of crude bio-oil production and wastewater treatment. The recent progress in advanced pyrolysis methods of seaweeds including catalytic pyrolysis, coprolysis, and catalytic co-pyrolysis for production of upgraded bio-oil was discussed. In addition, this work sheds light on the economic feasibility and environmental impacts of using seaweeds as a crude bio-oil feedstock.

## 2. Macroalgae/seaweeds

In a simple term, macroalgae are multicellular photosynthetic plant-like organisms that grow mainly in the seas and oceans. They play an important role as a valuable food to the seawater aquarium by providing a basic-chain food source for herbivorous animals. In addition, they work as a natural filter to aquarium system by reduction of total nitrogen and total phosphate levels and releasing oxygen. Based on their pigmentation variations, Kraan [37] categorized different seaweeds into three main phyla; Rhodophyta (red algae), Phaeophyta (brown algae) and Chlorophyta (green algae). Generally, Rhodophyta is the most species-rich macroalgal phylum containing about 6000 known species, followed by Chlorophyta and Phaeophyta with about 4500 and 2000 identified species, respectively [38]. Chlorophytes grow almost in all types of aquatic environments. However, Rhodophytes grow mostly in inter-tropical zones, while Phaeophytes grow mainly in temperate to cold water bodies [39]. Currently, seaweeds are cultivated for human food production, bio-fertilizers and hydrocolloids. China, Japan, Korea and Philippines account for about 72% of the global annual production of macroalgae [40]. Over  $23 \times 10^6$  tons dry biomass of macroalgae were produced during 2012 from aquaculture, which worth over US\$ 6 billion [41]. In recent years, macroalgal farming has been expanded fast

**Table 1**  
The most common biomass conversion routes showing the pros and cons of each method.

Conversion method	Description	Advantage	Disadvantage	Refs.
Fermentation	A microbial-assisted process for conversion of sugars into bioethanol	Low energy consumption High economy Mild reaction conditions	Long reaction time Difficult byproducts recycling	[12–14]
Anaerobic digestion	A microbial-assisted process for conversion of organic compounds into biogas	Low energy consumption Mild reaction conditions High economic feasibility	Long reaction time Generation of large amounts of digestate Usually requires a pretreatment step Compete with human food	[15,16]
Transesterification	A chemical process for biodiesel production from lipids rich in glycerides, mainly triacylglycerides (TAGs).	Fast process	Has negative environmental impacts Followed by transesterification which increases the production cost	[17–19]
Esterification	A chemical process for biodiesel production from lipids rich in free fatty acids such as waste cooking oil.	Fast process	Has negative environmental impacts	[17,20]
Pyrolysis	A thermal conversion process which is carried out at high temperature and oxygen-free environment. The products include bio-oil, non-condensable gas, and biochar.	Cost-effective and available feedstocks Fast conversion rate No additional pressure is required High energy products yield	High energy consumption Poor thermal stability High biofuel acidity Requires pretreatment for biomass drying	[21–25]
Liquefaction	A thermal conversion process which is carried out in the presence of solvents, high temperature and high pressure. The main energy products are bio-oil, non-condensable gas and biochar	No solvent is required Fast conversion rate High energy products yield No need for the drying step	High energy and solvent consumption Poor thermal stability High biofuel nitrogen content Huge amount of liquid byproducts	[26–30]
Gasification	A thermal conversion process that converts biomass into syngas under high temperature	Fast conversion rate High energy products yield No need for the pretreatment	Severe air pollution High energy consumption Low calorific value of the produced syngas	[31–33]

in different regions of the world from Southeastern Asia down to East Africa and South America due to the efforts to reduce the over-exploitation of natural resources and encouraged expanded seaweeds applications [42].

### 3. Chemical composition of macroalgae

There is a significant difference in the chemical composition of macroalgae comparing to the terrestrial plants and microalgae [40]. Table 2 shows a comparison between macroalgae and microalgae with regard to main chemical components and advantages/disadvantages of each group. In addition, Table 3 shows the main composition of some seaweeds in comparison to typical lignocellulosic biomass such as sawdust and rice husk. The main advantages of seaweeds over microalgae include the lower production cost due to ease harvest, and the well-established industrial infrastructure. Moreover, unique components such as carrageenan, laminarin, alginate, agar, mannitol, fucoidin, and fucose [38] make macroalgae different from lignocellulosic biomasses and even microalgae. Furthermore, the absence or low lignin content of macroalgae, as low as 0.03 g kg<sup>-1</sup> dw [38,43,44], offers ease processing and degradation without the costly pretreatment required in case of lignocelluloses [7].

Brown macroalgae are olive-greenish to dark brownish in color due to the abundance of the yellow-brown pigment fucoxanthin, which masks the chlorophyll's green color. This group of macroalgae includes the largest kelp (*Laminaria* sp.), which may reach 100 m in lengths at a growth rate as high as 50 cm day<sup>-1</sup> [59]. Kelps can be harvested from temperate and polar regions at depths below the low tide level, and are farmed extensively in China, Japan, and South Korea as food products [62]. Phaeophytes composition includes up to 55% of dry weight

laminarin and mannitol. Laminarin is a carbohydrate that can be hydrolyzed into glucose by laminarase (endo-1,3(4)-b-glucanase) [62]. Mannitol is a sugar alcohol that can be dehydrogenated into fructose, for further bioconversion into bioethanol [63]. In addition, phaeophytes contain cellulose and alginate, which are important structural polysaccharides providing mechanical strength to the cell wall.

Rhodophytes have a characteristic pink or red color due to the presence of phycoerythrin and phycoerythrin, which allow light capture and growth at relatively deep water. Therefore, red algae can be found in the subtidal and intertidal zones of the sea at 40 m water depths or, occasionally, as deep as 250 m [64]. Their composition varies according to the species but, generally, consist of cellulose, galactan, and glucan. The cell wall of red seaweeds contains two kinds of long-chain polysaccharides, namely agar and carrageenan, which are valued for gel-forming abilities and are used economically for thickening foods such as ice cream, yogurt, and pudding [65].

Due to their need for high light intensity, most of green macroalgae live at the shallowest water columns close to the surface. They are common in estuaries and bays where freshwater is mixed with salt water. Mostly, their composition includes cellulose and pectin as the main structural polysaccharide in the cell wall, in addition to starch as a food reserve [66]. However, the ash content and biochemical composition have wide seasonal variations due to the changes of environmental conditions. For instance, *Ulva* sp. recorded the highest carbohydrates value in June (61 dw%), while showed a gradual decline from 49 to 41 dw% during July to September, respectively [67]. Similarly, *Ulva intestinalis* showed the highest protein content of 27.7 dw% in winter, which reached to the lowest value of 6.7 dw% in spring [68]. Moreover, the wide seasonal variation in water characteristics leads to significant variations in seasonal areal biomass yields of seaweeds. In

**Table 2**  
Comparing the main biochemical composition, advantages and disadvantages of seaweeds with those of microalgae.

Algal species	Group	Proteins (dw%)	Carbohydrates (dw%)	Lipids (dw%)	Advantages	Disadvantages	Refs.
<i>Seaweeds</i>							
<i>Hypnea valentiae</i>	Rhodophyta	11.8–12.6	11.8–13.0	9.6–11.6	Wide distribution in natural ecosystems	Nuisance of the coastal areas	[24]
<i>Acanthophora spicifera</i>	Rhodophyta	12.0–13.2	11.6–13.2	10.0–12.0	Don't require agricultural land Easy to harvest	Lower growth rate than microalgae	[24]
<i>Laurencia papillosa</i>	Rhodophyta	11.8–12.9	12.0–13.3	8.9–10.8	High carbohydrate content	Contain high metal ions that are not desirable for bio-oil production	[24]
<i>Ulva lactuca</i>	Chlorophyta	11.4–12.6	11.6–13.2	9.6–11.4	A well-established industrial applications (e.g. agar, carrageenan, alginate, and colloids production)	Low lipid content	[45]
<i>Caulerpa racemosa</i>	Chlorophyta	11.8–12.5	16.0	9.0–10.5		Relatively lower CO <sub>2</sub> fixation rate	[46]
<i>Halimeda macroloba</i>	Chlorophyta	5.4	32.6	9.9			[47]
<i>Valoniopsis pachynema</i>	Chlorophyta	8.8	31.5	9.1			[48]
<i>Ulva reticulata</i>	Chlorophyta	12.8	16.9	8.5			[48]
<i>Enteromorpha compressa</i>	Chlorophyta	7.3	24.8	11.5			[49]
<i>Caulerpa cupressoides</i>	Chlorophyta	7.4	51.8	11.0			[50]
<i>Dictyopteris australis</i>	Phaeophyta	1.3	33.1	9.7			[51]
<i>Stoechospermum marginatum</i>	Phaeophyta	3.9	33.6	10.9			[51]
<i>Lyngaria stellata</i>	Phaeophyta	2.8	32.0	11.7			[24]
<i>Styopodium schimperi</i>	Phaeophyta	1.9	29.8	8.9			[24]
<i>Turbinaria turbinata</i>	Phaeophyta	0.2–2.9	20.5–33.9	8.0–13.8			[52]
<i>Microalgae</i>							
<i>Chlorella vulgaris</i>	Chlorophyta	52.0–56.4	17.3–19.2	12.4–15.7	Higher growth rate and biomass production	Difficult to harvest	[53]
<i>Chlorella</i> sp.	Chlorophyta	34.0–42.7	9.4–15.5	2.5–7.0		High energy consumption for processing	[51]
<i>Dunaliella tertiolecta</i>	Chlorophyta	61.3	21.7	2.9	Controllable cultivation conditions	Can be easily contaminated with undesired microbes	[54]
<i>Chlamydomonas reinhardtii</i>	Chlorophyta	61.7	3.3	12.19	Higher lipid content Higher CO <sub>2</sub> fixation rate	Used in many industrial applications as food supplement	[55]
<i>Chlamydomonas debaryana</i>	Chlorophyta	59.4	10.1	19.90		High cultivation cost	[55]
<i>Nannochloropsis</i> sp.	Ochrophyta	40.8	19.2	30.00	Higher efficiency of waste removal		[56]
<i>Nannochloropsis oculata</i>	Ochrophyta	24.0	6.9	14.46			[56]
<i>Spirulina platensis</i>	Cyanophyta	48.4–65.2	12.0–30.2	10.30–13.30			[57]
<i>Schizochytrium limacinum</i>	Heterokontophyta	51.0	24.0	14.00			[58]

Table 3

Proximate chemical composition of some representative macroalgae in comparison to lignocellulosic feedstocks.

Parameter	Macroalgae				Lignocelluloses	
	<i>Laminaria japonica</i>	<i>Sargassum fulvellum</i>	<i>Gelidium amansii</i>	<i>Ulva lactuca</i>	Sawdust	Rice husk
Group	Phaeophyta	Phaeophyta	Rhodophyta	Chlorophyta	Lignocellulosic wastes	
Ash (dw%)	31.5	46.0	8.6	18.9	5.9	15.4
Carbohydrates (dw%)	51.9	39.6	77.2	54.3	57.1	27.1
Lignin	0	0	0	0	28.7	46.6
Lipids (dw%)	1.8	1.4	1.1	6.2	–	1.2
Proteins (dw%)	14.8	13.0	13.1	20.6	0.7	2.9
Carbohydrate composition	Laminarin, mannitol, alginate, fucoidan, cellulose		Agar, carrageenan, cellulose	Starch, cellulose	Cellulose, hemicellulose	
References	[59]	[59]	[59]	[59]	[60]	[61]

addition, there is an interspecies variation within the same season and growth area. In a recent screening study, *Ulva intestinalis* showed the highest annual biomass yield of 61.5 g m<sup>-2</sup> year<sup>-1</sup>, while *Ectocarpus siliculosus* showed the lowest yield of 1.3 g m<sup>-2</sup> year<sup>-1</sup> [68]. Therefore, seaweed selection and the optimum harvest time for seaweeds should be determined based on the growth cycle, season and the desired end products.

#### 4. Algal wastewater treatment

Aquaculture farms and industrial effluents in many coastal areas around the world result in profound pollution and environmental degradation, which is considered as a serious problem in many countries. Treatment of the discharged water at the source is the most effective way to reduce pollution, however, most of factories and aquaculture farms do not have such *in-situ* treatment systems. In general, wastes originated from different sources can be classified into two main groups; biological wastes and non-biological wastes. The biological wastes are biodegradable and include those originated primarily from living resources, such as most of aquaculture wastes. Non-biological wastes include the recalcitrant pollutants such as chemical additives and heavy metals which are not easy to be degraded. Physical, chemical and biological treatment methods are the three available techniques to treat wastewater. Amongst, biological methods are the most advantageous in terms of low cost, simple operation and eco-friendly. Seaweeds can be used for biological removal of nitrogen and phosphorus in wastewater. They can take up nitrate and ammonia, which are the prevalent nitrogen compounds in domestic/urban sewage, agricultural and industrial water effluents. Devi and Gowri [69] reported a removal efficiency of 87.2% nitrate, 87.2% nitrite, 84.1% phosphate and 82.5% ammonia by *Enteromorpha flexuosa* from aquaculture farm discharge water after 20 days of cultivation. They also cultivated *Gracilaria verrucosa* and reported higher removal efficiency of 91.4%, 94.5%, 100%, and 99.3%, respectively. In addition, dissolved oxygen increased from 4.2 to 5.1 mg L<sup>-1</sup> and from 3.3 to 5.1 mg L<sup>-1</sup> with *E. flexuosa* and *G. verrucosa*, respectively. Interestingly, the growth of both seaweeds increased by 35.5% and 40.5%, respectively, using wastewater.

For recalcitrant pollutants and heavy metals removal from industrial wastewater, different technologies have been developed. Heavy metals can be precipitated into their hydroxide derivatives by addition of sodium hydroxide or lime (calcium hydroxide). This method is relatively cheap and can be used to remove bulk of heavy-metal ions. However, it cannot be used if a final clarification is required, and also have negative environmental concerns [70]. Metals can be removed from a solution using electrolysis in solid metallic form, with the advantages of no produced sludges and avoiding usage of extra chemical reagents. However, this method strongly depends on the energy price, and the final treatment cost depends on the amount of electricity consumed. Other methods, such as osmosis and reverse osmosis, membrane processes, dialysis, and electro dialysis, were recommended but they are tending to be used in very specialized applications [70].

Xiong et al. [71] reported advanced oxidation processes (AOPs) as one of effective technologies for waste removal, but their application is

limited for large-scale applications due to the required high maintenance and operational costs. Moreover, some byproducts with toxicity comparable to or greater than that of the initial compounds might be generated due to the incomplete mineralization of some chemicals during AOPs [72]. Purified oxidative enzymes, such as laccase, have been utilized to remediate recalcitrant pollutants. This technique is disadvantageous due to the low activities, high cost, and selectivity. Moreover, the elevated cost and limitations of the enzymatic reaction environment restrict the large-scales utilization of enzymes [73]. Application of adsorption processes for the removal of impurities from liquid or gaseous media are versatile and relatively simple. So far, some industrial important adsorbents such as silica gel, activated carbon, and alumina, have been developed with a porous surface structure which provides high surface area. However, the removal efficiency of the applied adsorbent is significantly affected by the background organic compounds [74]. Alternatively, biosorption is a kind of adsorption techniques which overcome the aforementioned disadvantages and, therefore, received increasing consideration over the last few years. It is a process by which soluble substances in a solution can be collected on a suitable interface of a living cell or organism. It is also used to describe the passive or physicochemical attachment of the substance to a biomass, thus excluding the metabolic or active-uptake processes.

Using bacteria and fungi for bioremediation has been widely investigated and is receiving a great attention nowadays. However, those microorganisms grow heterotrophically and require additional carbon sources for sufficient growth. Particularly, utilization of fungi in large-scale is limited because of the mycelial structure and slow catabolic kinetics [75]. Due to the autotrophic growth of algae, they are receiving a great attention in the field of bioremediation. Algal cell walls contain many polymers including cellulose, hemicellulose, pectin, and arabinogalactan proteins. The dominant functional groups including phosphoryl, carboxyl, and amines provide the algal cell walls with a negative charge, which activates the attraction of contaminants carrying cationic groups into the algal surface enhancing the biosorption process. Algal cells absorb the organic pollutants together with other growth nutrients into the cell through bioaccumulation. Latinwo et al. [76] evaluated the potential of collected green seaweeds to remove heavy metals from textile wastewater within 10–90 min. Results showed maximum removal efficiency for Fe, Ca, Mg, K, Ag and Cr by 87.5%, 99.9%, 59.7%, 57.2%, 100% and 86.8%, respectively, after 60 min. Ungureanu et al. [77] studied the biosorbents efficiency of *Sargassum muticum* and *Ascophyllum nodosum* for antimony (Sb(III)). Biosorption of Sb(III) by *S. muticum* was found to be a fast process with maximum biosorption capacities of 2.1 and 4.0 mg g<sup>-1</sup> at pH 2 and 7, respectively.

Navarro et al. [78] studied the biosorption of phenol by the marine seaweeds *Lessonia nigrescens* and *Macrocystis integrifolia*. Results showed the maximum adsorption efficiency of 35% at pH 10 using *Macrocystis integrifolia* due to a purely polar adsorption mechanism rather than an electrostatic adsorption. The study suggested that phenol was adsorbed onto the surface of seaweeds by formation of hydrogen bonds with the hydroxyl groups of the polysaccharides, such as alginates, that form the algal biomass structure. Using brown, green, and red seaweeds biomass for benzene and toluene biosorption, as two of the most soluble aromatic

hydrocarbons, was recently studied [79]. Results showed that phaeophytes have the highest removal efficiency for toluene and benzene (28 and 112 mg g<sup>-1</sup>, respectively). The biosorption mechanism was attributed to hydrophobic interaction mainly with lipids and, to a lesser degree, with proteins and carbohydrates by nonspecific Van der Waals interactions. In addition, micropollutants bioaccumulation by algae was confirmed as an important route for removal of trimethoprim, sulfamethoxazole, and triclosan [80]. Another advantage of biosorption is the intracellular biodegradation which is considered as the most effective method by which living algal cells can eliminate the chemical pollutants from the surrounding environment [71]. In that context, approximately 30–80% of recalcitrant chemicals such as carbamazepine, ibuprofen, tris(2-chloroethyl)phosphate, and caffeine in wastewater were degraded within the algal cells [81–83]. Thus, biosorbents activity of seaweeds could be considered as a potential alternative to diminish the toxic effect of different pollutants in the aquatic ecosystems, which provides a promising approach for integrated energy production.

## 5. Thermal conversion of seaweeds

The process of biomass conversion aims to generate energy by converting biological materials to condensed energy product such as biogas, biodiesel, bioethanol, or crude bio-oil. Among different biomass conversion methods, thermal conversion has recently gained a great attention because it is much faster than biological processes, such as anaerobic digestion or fermentation. Due to low lipids content of seaweeds, thermochemical conversion may be more suitable than biochemical techniques such as lipid transesterification [84]. Kan et al. [85] concluded that thermochemical processes can convert not only lipids, but also other organic components such as proteins and carbohydrates into liquid and gaseous fuels. Therefore, thermal conversion of nuisance wild macroalgae for bioenergy production is a promising approach for renewable fuel production and environmental improvement. In addition, thermal decomposition can be used for energy production from some wastes that are biologically undegradable, such as plastic [86]. Thermal conversion processes mainly include pyrolysis, hydrothermal liquefaction, and gasification, in addition to direct combustion for heat generation (Fig. 1). Although all these methods have the same basic mechanism where heat is used to convert biomass into usable energy compounds, the amount of air supply and the energy output are quite different. For example, direct combustion needs excessive oxygen to produce energy in the form of heat, while pyrolysis takes place in the absence of air to produce mainly crude bio-oil [87]. Among different methods, pyrolysis is receiving more attention due to the wide range of

feedstocks, higher conversion efficiency and desirable end products with high yield of liquid bio-oil [88–90]. However, a pretreatment step is required to improve the conversion of residual biomass to bioenergy.

### 5.1. Pretreatment

For pyrolysis, macroalgal biomass must be dried to maintain a stable and efficient conversion process as well as oxygen-free conditions. Approximately, 0.7 MJ kg<sup>-1</sup> is the energy required for macroalgal biomass drying with a moisture content of 88%, which was reported to be higher than the corresponding lower heating value (LHV) of the dried seaweeds [91]. In addition, storage of high moisture-contained biomass for long time results in biomass deterioration and energy loss [92]. Therefore, dewatering of wet seaweeds to 20–30% allows lower energy consumption during drying and prevents the spoilage of the biomass [93]. Application of electro-osmotic dewatering of seaweeds (Fig. 2A) before drying was reported as a useful technique to reduce the energy requirements of the drying process [94]. In addition, on-site mechanical dewatering methods; e.g. pressing and centrifugation; reduce the energy cost required for biomass transportation. Mobile compressors have already been used for on-site seaweed dewatering (Fig. 2B). After dewatering, fuel-fired ovens may be used for drying and torrefaction of the biomass, however, the cost of the overall process increases greatly with undesired emissions. Alternatively, sun drying is the most commonly eco-friendly used method since it depends on the solar radiation and, therefore, reduces the drying energy costs. However, it is weather dependent and requires large areas as only around 100 g d<sup>-1</sup> m<sup>-2</sup> of the dried biomass can be produced [95]. Consequently, the

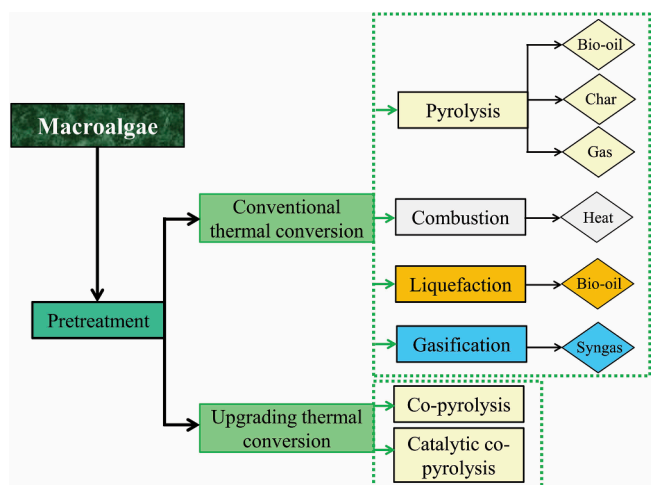


Fig. 1. Different thermochemical conversion routes of macroalgal biomass.

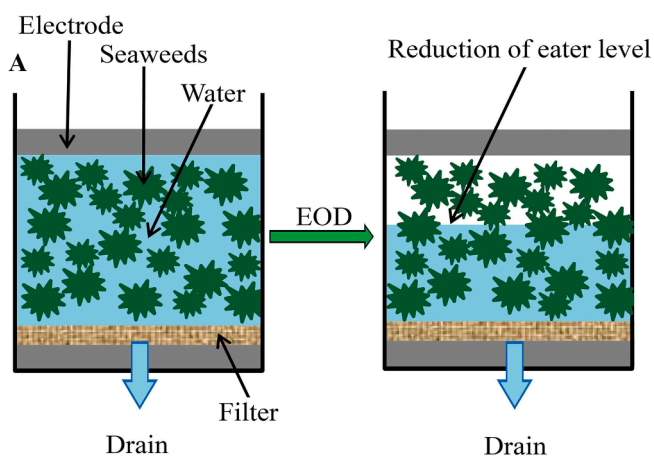


Fig. 2. Concept of electro-osmotic dewatering (EOD, A), and on-site mechanical dewatering of seaweeds at Qingdao, Shandong Province, east coast of China (B).

feasibility of seaweeds-to-fuels relies on finding a more cost-effective and controllable dewatering/drying method.

In addition to the high water content, macroalgae have significantly higher amounts of inorganic compounds, mainly metal ions such as K, Na, Mg and Ca, than lignocellulosic biomasses, causing fouling problems during thermal processing such as ash fouling and agglomeration in fluidized-bed reactor [96–98]. Generally, metal ions will also be attached on carbon surface rather than be evaporated during thermal conversion [99]. Therefore, removal of these inorganic elements improves the conversion process and upgrades the produced bio-oil [98,100,101]. In addition, Choi et al. [102] concluded that removal of macroalgae-specific organic components including alginate, laminarin, and mannitol is required in order to increase and upgrade the conversion products. In that regard, using seaweed residues after extraction of valuable industrial compounds provides a cost-effective enhanced energy recovery from wastes. Consequently, different pre-treatments using acid, water or  $\text{CaCl}_2$  have been studied to remove the undesired inorganic and organic components from seaweeds.

Ross et al. [97] carried out the pretreatments of seaweeds *Fucus vesiculosus*, *Laminaria hyperborea* and *Macrocystis pyrifera* by water and weak acids. They found that Mg, K and Na in all studied algae were reduced by 30–40% after water pretreatment, with insignificant changes in Ca content. However, acid pretreatment resulted in significant reduction in all metal ions by over 90%. Ly et al. [101] reported that inorganic content of *Cladophora socialis* decreases significantly from 19.3 to 9.2 dw% after acid washing. In addition, the removal rates of Ca, Mg, P, Al, K, and Na were 90.5, 92.7, 74.3, 97.6, 97.6, and 97.9%, respectively. Hu et al. [50] confirmed that  $\text{H}_2\text{SO}_4$  was more effective on the removal of metal ions than other tested acids and water. In addition to removal of inorganic compounds, acid pretreatment showed significant removal of organic components from biomass [50,103,104]. However, washing with water recorded a relatively slight reduction in some organic compounds, with insignificant changes in fucoidan, laminarin and alginic acid contents of seaweeds biomass [97]. Furthermore, Bae et al. [105] confirmed that acid pretreatment significantly reduces the ash content during pyrolysis of *Undaria pinnatifida*, which enhanced the bio-oil yield by 15%. Pretreatment using  $\text{CaCl}_2$  was also utilized to soften the macroalgal cellular structures for extraction of the valuable fucoidan [106,107]. Due to softening the cellular structures and the efficient removal of catalytic minerals,  $\text{CaCl}_2$  pretreatments increased the fatty acids ratio and reduced sugar derivatives in the produced heavy oil; with reduction of anhydrous dimers of mannitol and sorbitol in the light oil [102]. Recently, pyrolysis products of *Enteromorpha clathrata* washed with 7% phosphoric acid, sulfuric acid, and hydrochloric acid were studied [98]. Results showed that washing significantly increased the yield of bio-oil in favor of biochar, with higher aliphatic hydrocarbons contents in the bio-oil. Thus, washing of seaweeds as a pretreatment plays a key role to enhance the bio-oil yield and significantly influences its characteristics. Not only the bio-oil quality was improved as a result of pretreatment, but also the characteristics of the biochar as a solid fuel source were improved. For example, the biochar produced from acid-pretreated *Saccharina japonica* showed relatively higher carbon content and higher heating value (HHV) with lower ash content [102]. Although other chemicals were used successfully for the pretreatment, e.g. ethanol, it is not suitable for large commercial scale due to the high processing cost.

## 5.2. Pyrolysis

Among different thermal conversion processes, pyrolysis is considered as the most efficient method with highest fuel-to-feed ratios [88]. It is a highly complex process that involves many thermochemical reactions and physical transformations, which has been widely investigated using thermogravimetric analysis (TGA) under both non-isothermal and isothermal conditions [108–110]. It occurs usually at 400–700 °C in the absence of oxygen resulting in three final products;

bio-oil, biochar and non-condensable gas consisted mainly of  $\text{H}_2$  and  $\text{CO}_2$  [111,112] as shown in Table 4. The liquid bio-oil obtained from pyrolysis is easier to be stored and transported in comparison to syngas produced by gasification or heat produced by direct combustion. In addition, the gas produced from pyrolysis showed net calorific value of 10–20  $\text{MJ Nm}^{-3}$ , which is higher than the syngas produced from gasification and combustion (4–15  $\text{MJ Nm}^{-3}$ ), which is attributed to anaerobic conditions during pyrolysis. Moreover, less dioxin and trace heavy metals emissions are produced in the gas stream of pyrolysis because it can be achieved at lower temperatures compared to combustion and gasification [113]. Nevertheless, the crude bio-oil from pyrolysis is typically inappropriate for direct use in engines because of its high viscosity and low pH, which might result in severe engine deposition and corrosion [114]. In addition to bio-oil uses, biochar and gas have high economic value. Moreno-Piraján et al. [115] recorded the high adsorption capacity of biochar produced from cow bone residues for the heavy metals  $\text{Cu}^{2+}$  and  $\text{Pb}^{2+}$ . Mullen et al. [116] also reported the effective use of biochar for metal removal from wastewater, with adsorption efficiency up to 50 and 80% for  $\text{Cu}^{2+}$  and  $\text{Pb}^{2+}$ , respectively. In addition, biochar can be used for soil improvement [117], as renewable solid fuel [118,119] or for enhanced energy recovery from biomass [120,121]. Concerning the produced non-condensable gases, they can be re-circulated to the process and thus serve as an additional heating source [122,123].

Recently, there is a growing interest in bio-oil and renewable carbonaceous materials production from seaweeds [98,132–134]. Pyrolysis, however, requires relatively dry materials as discussed in section 8.1, and, therefore, it is only feasible after extraction of high-value products from macroalgae. Thermochemical behavior of various dry marine macroalgae differs greatly, not only from the terrestrial biomasses, but also between each other [135,136]. Different kinetic parameters of pyrolysis process can be calculated from the characteristic parameters obtained by differential thermogravimetric (DTG) and thermogravimetric (TG) curves during TGA analysis. Li et al. [137] investigated pyrolysis of the brown macroalga *Sargassum pallidum* using TGA and recorded average activation energy of 203.5 and 202.9  $\text{kJ mol}^{-1}$  using Kissinger-Akahira-Sunose and Flynn-Wall-Ozawa methods, respectively. Recently, Ali and Bahadar [110] used four iso-conversional kinetic methods to study the degradation kinetics of *Sargassum* sp. and found an exponential increase in the apparent activation energy of pyrolysis from 35 to 640  $\text{kJ mol}^{-1}$  by increasing the thermal degradation from 10% to 90%.

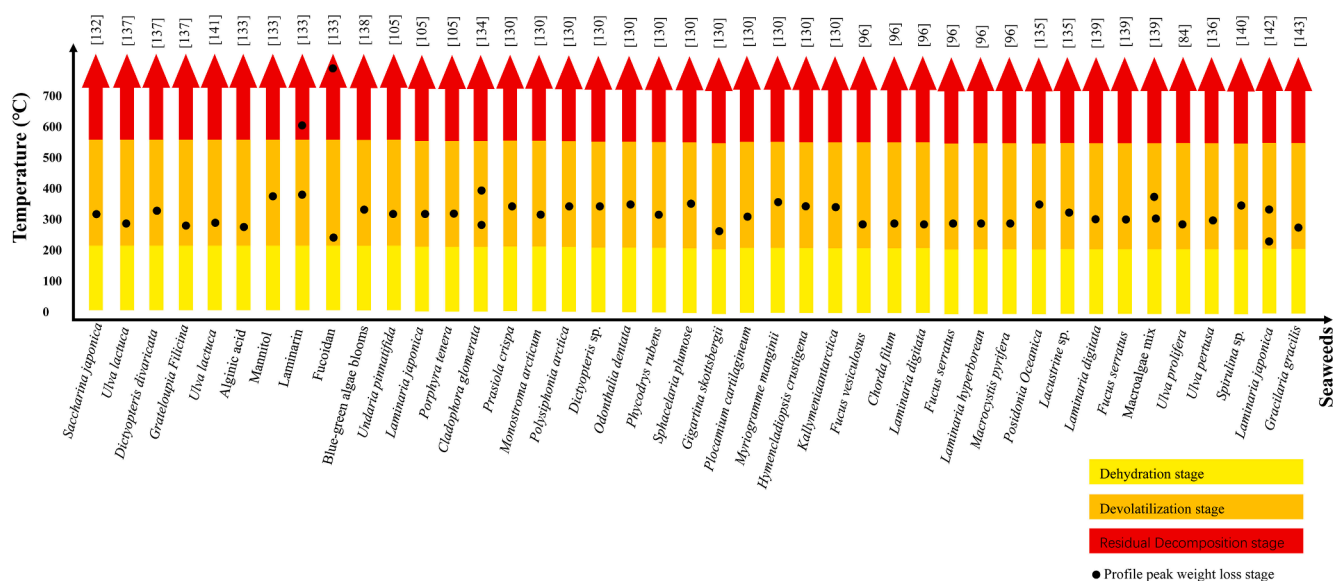
Generally, there are three main stages of seaweed thermal decomposition; dehydration (stage I), devolatilization (stage II) and decomposition of carbonaceous solids (stage III) [138,139]. Fig. 3 represents a comparison of the thermal degradation stages of different seaweeds at different decomposition temperatures with the profile peak of weight loss. The first thermal degradation stage occurs within a temperature range up to 200 °C. It involves a relatively small weight loss, which is attributed to the evaporation/removal of moisture and decomposition of light volatile compounds. Further, a greater mass loss can be recorded within the temperature range of 200–550 °C as a result of devolatilization of carbohydrates, proteins and lipids (the second stage). The third stage involves a slow weight loss that usually continues, in most cases, up to 600 °C due to gradual loss of volatile metals and decomposition of carbonates at low rates. Thus, the maximum weight loss stage of the pyrolysis profile of seaweeds occurs during the second stage of decomposition.

Apart from thermal treatments of macroalgae by TGA, identification of the main chemical compounds of the bio-oils from different macroalgae has been grossly investigated [46,140,141]. Fig. 4 represents a comparison of elemental compositions of bio-oil produced from pyrolysis of different seaweeds. Generally, seaweed bio-oil mainly contains carbon (45.3–68.4%) and oxygen (12.9–43.6%). However, hydrogen and nitrogen represent relatively much lower proportions, with very little or absence of sulfur.

**Table 4**  
Yields and characteristics of pyrolysis products from different macroalgae.

Seaweeds	Product yield (%dw)			BTP	Characteristics of bio-oil				References
	Bio-oil	Biochar	Gas		HHV	Flash point (°C)	M	D	
<i>Enteromorpha clathrata</i>	41.20	41.50	17.3	500	12.01	NA	NA	NA	[124]
<i>Sargassum natans</i>	33.70	47.90	18.4	500	8.68	NA	NA	NA	[124]
<i>Saccharina japonica</i>	48.40	32.30	NA	450	28.70	84.0	1.49	NA	[125]
<i>Saccharina japonica</i>	44.99	34.20	20.81	350	24.80	NA	6.90	NA	[101]
<i>Ulva lactuca</i>	65.00	20.00	NA	550	25.70	NA	26.60	0.98	[126]
<i>Undaria pinnatifida</i>	39.50	60.30	30.0	500	23.33	NA	NA	NA	[105]
<i>Laminaria japonica</i>	37.50	54.20	34.0	500	33.57	NA	NA	NA	[105]
<i>Porphyra tenera</i>	47.40	60.00	22.0	500	29.74	NA	NA	NA	[105]
Algal seaweed meal	35.60	30.80	33.7	550	26.21	NA	NA	NA	[127]
<i>Cladophora glomerata</i>	39.00	40.00	22.0	500	17.19	NA	NA	NA	[128]
<i>Posidonia oceanica</i>	52.40	22.80	24.8	500	26.10	NA	12.68	1.25	[129]
<i>Laminaria digitata</i>	32.00	34.00	32.0	500	23.08	NA	4.60	NA	[130]
<i>Fucus serratus</i>	37.00	42.00	23.0	500	32.46	NA	1.20	NA	[130]
<i>Laminaria japonica</i>	35.4	29.0	NA	500	29.60	NA	70.7	NA	[131]
Macroalgae mix	37.00	37.00	30.0	500	26.40	NA	3.70	NA	[130]

NA Not available; BTP Optimum temperature (°C) for maximum production; HHV Higher heating value of bio-oil (MJ kg<sup>-1</sup>); D Density at 40 °C (g cm<sup>-3</sup>); M Moisture content (%).



**Fig. 3.** Comparison of the main stages of thermal degradation of different seaweeds using TGA/DTG pyrolysis.

In addition, the yield and characteristics of pyrolysis outputs vary in different macroalgae. Bio-oil yield of seaweeds ranges from 32 to 65 dw %; the yield of biochar is between 20 and 60 dw%; while that of the gaseous product is within the range of 17 to 34 dw% (Table 4). However, bio-oil from different seaweeds recorded relatively HHV (up to 33.57 MJ kg<sup>-1</sup>) compared to that of lignocellulosic biomasses, which is more desirable from energy aspect. The main recorded chemical compounds in different seaweed bio-oils have been studied by many researchers. Table S1 (Supplementary data) shows a survey for the main chemical compounds in pyrolytic bio-oils of various seaweeds. The major chemical compounds in the bio-oils are similar to some extent, which might be attributed to the interspecies chemical composition similarities of macroalgae. The main components of bio-oils are hydrocarbons, aldehydes, ketones, alcohols, carboxylic acids, phenols, and their derivatives, with minor proportions of heterocyclic compounds such as pyridines, furans, and pyrans (Table S1, Supplementary data).

Several studies have compared the macroalgal pyrolysis with that of lignocellulosic biomasses. Trinh et al. [126] compared energy recovery and different products from pyrolysis of macroalgae, lignin, wood and straw. Macroalgae produced 65 dw% bio-oil with 76% bio-oil energy recovery, while lignin yielded 47 dw% bio-oil with 45% bio-oil energy

recovery. The HHV of wood, straw, lignin, and algae bio-oils were 24.0, 23.7, 29.7, and 25.7 MJ kg<sup>-1</sup> dry weight, respectively. As a general statement, lignocellulosic biomasses require higher pyrolysis temperatures than macroalgae, likely due to the cellulose, hemicellulose, and lignin components [139,142]. Contrary to lignocellulosic biomasses such as husks, straws and sawdust, seaweeds have lower thermal stability and, therefore, they volatilize at lower temperatures. In a previous study, Kebelmann et al. [138] pyrolyzed psychrophilic macroalgae collected from the Arctic region and recorded that the maximum decomposition rate of *Prasiola crispa* was within the temperature range of 220–320 °C, which was lower than that of lignocellulosic biomasses. Wang et al. [143] recorded the release of volatiles during pyrolysis of seaweeds earlier than that of lignocelluloses. In addition, seaweeds pyrolysis was exothermic resulting in heat, meaning that less net energy input is required [143]. Ross et al. [96] compared the thermal degradation behavior of five brown seaweeds with that of terrestrial plants. Results showed a lower proportion of phenolic compounds in macroalgae. As shown in Table S1, seaweed pyrolysis showed almost the absence of some phenolic fragments common in lignocellulosic bio-oil such as methoxyphenols. Absence of such phenolic fragments is attributed to the absence of lignin in seaweeds [96,144]. This is also

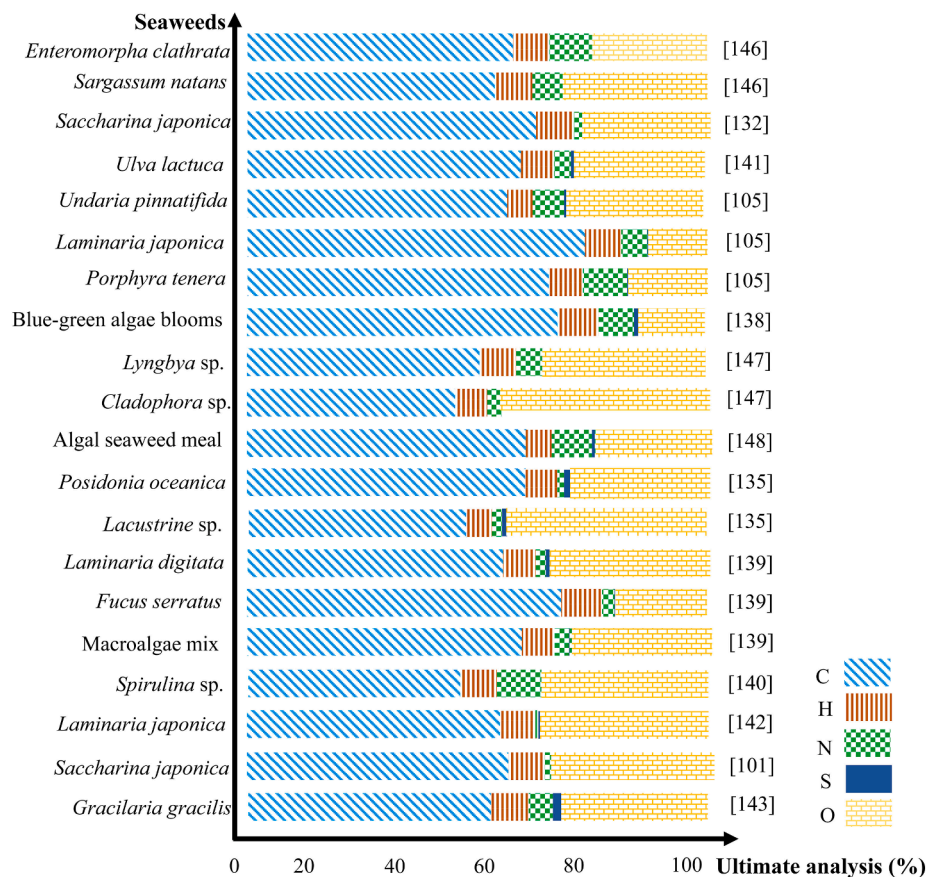


Fig. 4. Elemental compositions of bio-oil produced from pyrolysis of different seaweeds.

advantageous for higher bio-oil quality, because the presence of such phenolic compounds results in a difficulty during deoxygenation process

Table 5

Recent progress on advanced pyrolysis methods of seaweeds (catalytic pyrolysis, co-pyrolysis, and catalytic co-pyrolysis).

Seaweeds/components	Co-feeding element of high C & H contents	Pyrolysis system, conditions, and applied catalyst	Results	Refs.
<i>Enteromorpha prolifera</i> (EP)	Rice husk (RH)	Fixed-bed reactor, 400–600 °C; 5–25 °C min <sup>-1</sup> ; N <sub>2</sub> gas; 100 mL min <sup>-1</sup>	Results revealed positive synergy existed between EP and RH which increased the oil yields and improved the oil quality.	[139]
<i>Enteromorpha clathrata</i> polysaccharides and <i>Sargassum fusiform</i> polysaccharides	Cellulose	TGA instrument coupled with GC/MS; N <sub>2</sub> gas; 100 L min <sup>-1</sup>	Results also revealed the existence of synergy & the simulation results were consistent with the experimental results.	[149]
<i>Enteromorpha clathrata</i> polysaccharides and <i>Sargassum fusiform</i> polysaccharides	Cellulose	Stainless steel fixed bed reactor; 550 °C; ZSM-5 catalyst; N <sub>2</sub> gas; 0.8 L min <sup>-1</sup>	Results showed an increase in oil yields due to ZSM-5 and a great reduce in acids and N-containing compounds in the oil, while furans and ketones were greatly increased.	[150]
<i>Laminaria japonica</i>	Polypropylene	Batch-type fixed-bed reactor; TGA instrument; Py-GC/MS; N <sub>2</sub> gas; 50 mL min <sup>-1</sup> ; 500 °C; Catalysts used are: HZSM-5, HBETA and HY	Results revealed a sharp decrease in the oil water content and improvement in oil quality due to increased mono-aromatic hydrocarbons in the oil.	[151]
<i>Enteromorpha clathrata</i>	Rice husk (RH)	Fixed bed reactor; 190, 320 and 550 °C; N <sub>2</sub> gas; 200 mL min <sup>-1</sup>	Results showed positive synergistic effects, and a release of N- substances was inhibited by addition of RH at low temperature (<190 °C) range.	[147]
<i>Laminaria japonica</i>	Polypropylene	U-type quartz fixed-bed reactor; TGA instrument; Py-GC/MS; N <sub>2</sub> gas; 50 mL min <sup>-1</sup> ; 500 °C; Catalysts used are: HZSM-5, mesoporous MFI, Pt/mesoporous MFI, & mesoporous Al-SBA-16.	Result showed a decrease in oxygenates, acids, and wax components, and increase in hydrocarbons (aromatics and light).	[148]
<i>Ulva lactuca</i>	–	Quartz tubular reactor; N <sub>2</sub> gas; 100 mL min <sup>-1</sup> ; 460 °C; HZSM-5 catalyst	Results indicated a huge amount of aromatic hydrocarbons and high denitrogenation effect for <u>amides</u> , <u>amines</u> and <u>nitriles</u> , however, with a great amount of coke.	[152]
<i>Laminaria japonica</i>	–	Py-GC/MS analyzer; Catalysts used are: nanoporous Al-MCM-48 and hierarchical Meso-MFI zeolite	Meso-MFI exhibited a higher activity in deoxygenation and aromatization than Al-MCM-48, producing higher yields of aromatics, gases and phenolics owing to the strong acidic sites.	[153]



which requires further bio-oil upgrading.

### 5.3. Upgraded bio-oil

Algae, most especially seaweeds, have been acknowledged by many researchers as a promising future energy source due to the high photosynthetic efficiency and biomass productivity [2,18,110,136,145,146]. However, bio-oil produced from pyrolysis of seaweeds often exhibits severe instability as a result of high oxygen contents. In addition, nitrogen-containing compounds and sulphur should be as minimum as possible to reduce the NO<sub>x</sub> and SO<sub>x</sub> emissions. Thus, it requires either direct catalytic reforming or upgrading via co-pyrolysis, with or without catalysts, to improve the bio-oil stability and emission performance [25,147,148]. Many studies have been carried out on catalytic pyrolysis of lignocelluloses and other organic wastes, indicating that application of a suitable catalyst improves the heating value and lowers the oxygen and nitrogen contents of the bio-oils. However, studies on catalytic pyrolysis of seaweeds are still very limited compared to those of terrestrial biomass and wastes, while investigations on co-pyrolysis of seaweeds are very few in literature. It is thus imperative to focus more research attention on the advanced pyrolysis methods of seaweeds via application of different catalysts or by employment of several upgrading methods such as catalytic co-pyrolysis in order to properly evaluate the pyrolysis of seaweeds as a potentials and effective energy source. This section of the article, therefore, represents a holistic summary of the recent progress in advanced pyrolysis methods of seaweeds (catalytic pyrolysis, co-pyrolysis, and catalytic co-pyrolysis), and optimization of the co-pyrolysis parameters/conditions towards enhanced bio-oil production. Table 5 summarizes the recent progress on the most commonly advanced pyrolysis methods of seaweeds.

#### 5.3.1. Catalytic pyrolysis of seaweeds

Generally, biomass catalytic pyrolysis involves the application of a suitable catalyst during thermochemical decomposition process, which is irreversible reaction. The products of biomass catalytic pyrolysis often differ owing to the utilized catalyst type (e.g., zeolites, solid phosphoric acid, metal loaded catalysts, etc.). The microporous zeolites, e.g., HZSM-5 among other types of catalysts, are active for production of hydrocarbons [154–156]. However, the transformation pyrolyzates of large molecular sizes from biomass pyrolysis over microporous zeolites is limited due to the difficulty with which the pyrolysis products of large particles could diffuse into the microporous zeolites' small pores. Alternatively, mesoporous catalysts, e.g., SBA-15 and MCM-41, can be used for active conversion of pyrolyzates of large molecules [157–159]. Besides, deoxygenation reactions take place during catalytic pyrolysis and result in a significant decrease in the bio-oil oxygen content, with an increased heating value and removal of acids. However, the bio-oil from catalytic pyrolysis of biomass, including that of algae, contains considerable amounts of coke and low oxygenates, but with improved miscibility, most especially, with the petroleum-derived liquid fuels.

Research results revealed that lignocelluloses have been widely employed as catalytic pyrolysis feedstock compared to macroalgae, which are currently receiving a considerable attention as a new feedstock for crude bio-oil production. For example, Lorenzetti et al. [152] performed catalytic pyrolysis of seaweeds over HZSM-5 catalyst. They observed a huge proportion of aromatic hydrocarbons and high denitrogenation effect for amides, amines and nitriles, however, with a great amount of coke. Lee et al. [153] also carried out catalytic pyrolysis of the seaweed *Laminaria japonica* over a nanoporous Al-MCM-48 and hierarchical Meso-MFI zeolite (Meso-MFI) via direct Py-GC/MS to examine the impact of different catalysts on the products distribution and chemical compositions of the produced bio-oil. Results showed that Meso-MFI exhibits higher activity in aromatization and deoxygenation compared to Al-MCM-48, resulting in higher yields of aromatics, gases and phenolics due to the strong acidic sites which accelerate the cracking of pyrolysis oil molecules. The production of large amount of coke has been

identified by several researchers as a major challenge to the catalytic pyrolysis. Thus, the quest for a lasting solution to this problem has recently paved way for the employment of other biomass thermal conversion methods such as co-pyrolysis with different feedstocks.

#### 5.3.2. Co-pyrolysis of seaweeds

Recent progress in pyrolysis of seaweeds focused on application or development of advanced pyrolysis technologies, known as co-pyrolysis and catalytic co-pyrolysis, with optimization of the co-pyrolysis parameters/conditions for enhanced bio-oil production. Co-pyrolysis refers to the pyrolysis of two or more feedstocks to enhance the products yield and quality via a synergistic effect of different intermediates during the reaction [21,47,160,161]. Thus, ideal co-pyrolysis of seaweeds involves pyrolysis with a feedstock which contains a lot of hydrogen and carbon such as plastics, tires or rubbers. Co-pyrolysis of biomass with plastic materials significantly enhance the hydrogen and carbon contents of the produced bio-oil, resulting in improved fuel quality at higher bio-oil yields [111,152,162,163]. Likewise, comprehensive reviews on the different aspects of pyrolysis technology of different biomass feedstocks and polymers have been presented by many researchers. Zhang et al. [164] carried out a comprehensive review on pyrolysis technology of polymers with lignocellulosic biomass, particularly. However, their focus was on the catalytic co-pyrolysis chemistry. Uzoejinwa et al. [111] recently provided a holistic and comprehensive review on the recent advances, findings and perspectives of seaweeds co-pyrolysis technology with different plastic wastes for enhanced bio-oil production. However, this section of the present article specifically represents the current advances in research and development in co-pyrolysis of seaweeds or their components and various feedstock materials containing relatively high carbon and hydrogen contents, with or without catalysts.

Detailed information on co-pyrolysis of seaweeds with other feedstock materials containing lots of carbon and hydrogen towards enhanced biofuels production is currently not readily available in literature. For the first time, Lee et al. [151] investigated the pyrolysis and co-pyrolysis of the brown seaweed *Laminaria japonica* and polypropylene over a mesoporous material Al-SBA-15 using a fixed-bed reactor and Py-GC/MS (Table 5). Results showed that co-pyrolysis of *Laminaria japonica* with polypropylene results in significant reduction in the water content of the bio-oil and improves the bio-oil quality as the mono-aromatic hydrocarbons increased owing to catalytic co-pyrolysis. Recently, Uzoejinwa et al. [139] investigated the co-pyrolysis of seaweeds with lignocellulosic biomass and optimized the production of the pyrolysis products. Authors confirmed a positive interactive synergetic effect between lignocellulosic biomass (rice husk) and seaweeds (*Enteromorpha prolifera*) during co-pyrolysis, which resulted in significant increase in the bio-oil yield and concurrently improved the products quality. Wang et al. [149] also studied the co-pyrolysis mechanism of seaweed polysaccharides and cellulose through macroscopic analysis and molecular simulations to investigate the synergistic effects. The study confirmed the existence of synergy during co-pyrolysis, and the simulation results were found to be consistent with the experimental results. According to Brebu et al. [165], synergetic formation of aromatic hydrocarbons is the main target of co-pyrolysis. Generally, synergetic effect represents the interactions that exist between two or more elements which results in a total effect that is higher than the integral sum of the effects from the individual feeding elements, which usually improves the quantity and quality of the produced bio-oil, or might also worsen the characteristics of the resultant products [139]. Kim et al. [148] carried out the co-pyrolysis of *Laminaria japonica* and polypropylene using fixed-bed reactor and Py-GC/MS over different catalysts. They observed the reduction of acids, oxygenates, and wax contents of components via catalytic upgrading, while hydrocarbons content was considerably increased, enhancing the quality and economic value of the bio-oil. Likewise, Xu et al. [147] studied the synergetic mechanism during co-pyrolysis of seaweeds and rice husk through char/coke characteristics investigation. Their results confirmed

synergistic effects of co-pyrolysis which resulted in significant inhibition in the released nitrogenous substances at lower temperature (<190 °C). Catalytic co-pyrolysis of seaweeds polysaccharides with cellulose over ZSM-5 did not only increase the yields of bio-oil, but also significantly reduced the acids and nitrogen-containing compounds, while furans and ketones were considerably increased, suggesting the catalytic co-pyrolysis as a better method over co-pyrolysis [150]. Thus, the use of a suitable catalyst in the co-pyrolysis of seaweeds is one of the most recent areas of interest, where much attention is currently focused on high quality bio-oil production.

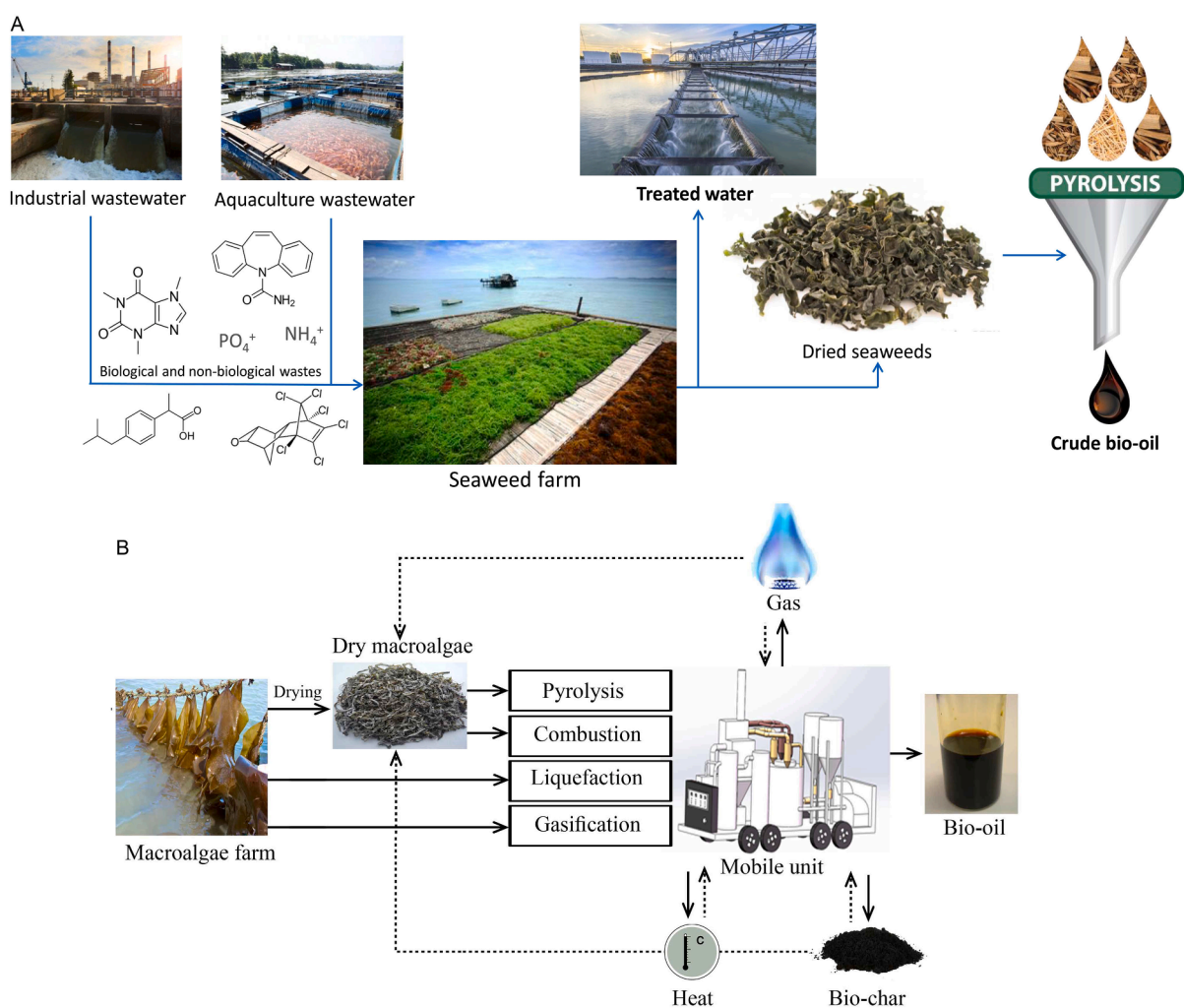
Likewise, optimization of biomass co-pyrolysis parameters/conditions such as reaction temperature, heating rate, residence time, feed-stock blending ratio, inert gas flow rate, particle size, and feed-to-catalyst weight ratio is another interesting aspect that is recently gaining increasing attention. For example, Hu et al. [166] investigated the co-pyrolysis of petroleum sludge and waste biomass using a response surface methodology to evaluate the interaction effects of temperature and heating rate on bio-oil and biochar yields. The study revealed a significant interaction between heating rate and sawdust percentage and between the heating rate and temperature on the bio-oil yield. However, several studies on optimization of individual pyrolysis of biomass have been carried out by many researchers [167–169], but for the first time, Uzoejinwa et al. [139] performed an optimization study on the co-pyrolysis of seaweed biomass. They studied the interaction effects of three effective parameters during seaweed co-pyrolysis on the yield of

bio-oil and biochar, then performed a simulation and modeling analysis in order to predict the optimal conditions to maximize the bio-oil yield. The study concluded that the synergistic effect between seaweeds and rice husk during co-pyrolysis resulted in significant increase in the products' yields and improved the co-pyrolysis products' quality. Moreover, co-pyrolysis of seaweeds and lignocellulosic biomasses was reported to improve the biochar properties [23].

## 6. Environmental impacts and economic feasibility

### 6.1. Environmental impacts

Environmental issues represent one of the key parameters used to evaluate a process acceptability to drive the next generation of a certain economic opportunity. Seaweeds grown in different kinds of wastewater discharges can provide an advantageous route for water treatment and biodegradation of recalcitrant compounds at comparatively lower cost and reduced ecological risks. While constructing such integrated system, achieving the maximum biodegradation/biosorption rate with the minimum cost is a critical issue for large-scale application. In addition, some concerns can arise about the utilization of seaweeds for human or animal feed due to the high pollutants within the biomass. Therefore, more investigations and efforts are required in order to overcome these concerns. Although using microbial consortium, consisted from microalgae and bacteria, for biological processes enhanced the process



**Fig. 5.** Integrated concept of seaweeds grown in wastewater for crude bio-oil production and wastewater treatment (A) and pathways of thermochemical conversion of seaweeds to bio-oil using on-site thermal decomposition (B), dotted lines represent heat recovery.

efficiency than the individual system, integration of algal biotechnologies with other current technologies have rarely been studied. Most of the recent studies on application of algae-based technologies are devoted and focused on laboratory-scale using individual methods, and only few studies have been conducted on pilot- or large-scales. Thus, thermal decomposition of seaweeds grown in wastewater for integrated crude bio-oil production and water treatment provides the best route for biomass utilization (Fig. 5A). However, more studies should be conducted to investigate the dual use of seaweeds for efficient removal of recalcitrant compounds and their impact on the produced bio-oil.

Nowadays, CO<sub>2</sub> emissions are discussed as an international political issue, and CO<sub>2</sub>-induced global warming is in the everyday news. Due to the high economic cost of CO<sub>2</sub> emissions reductions, biomass-generated fuels are receiving increasing attention as more economically and eco-friendly alternatives to the conventional petroleum fuel [20,170,171]. In addition to bioenergy production, macroalgae have a great potential for CO<sub>2</sub> bioremediation without competing with terrestrial crops for farm land or freshwater [24,172–174]. Previous work has shown that production of one ton of dry seaweeds approximately absorbs 960 kg net CO<sub>2</sub> [175]. Moreover, seaweeds can be grown in seawater without additional nutrients or pesticides [40,176], improving the water quality in which it is grown [177,178]. Therefore, macroalgal biomass represents a promising biofuel feedstock to provide environmentally-feasible alternatives for fossil fuel.

On the other hand, application of commercial macroalgal bioenergy systems expands the usage of algae farming, and thus special attention should be paid to their prospective impacts on coastal and marine environments. Such impacts include nutrient depletion, decrease of biodiversity, possible alteration of natural habitats, change of hydrology, and coral reefs disturbance [59]. Currently, environmental impacts of macroalgae farming might be seen as a minor issue in some cases, and may even have some benefits to increase the populations of invertebrates and fish in the area where seaweeds grow [179]. However, extensive long-term wild-harvesting of seaweeds will affect the environmental sustainability by disturbing the wild life and biodiversity of sea ecology [180]. Environment & Heritage Service statement [181] indicated that the depletion of *Laminaria digitata* in France might be attributed to different factors including over-exploitation. Therefore, sustainable harvest should be considered, using advanced instruments that leave parts of the vegetative organs of seaweeds for re-growth. Overall, a balance must be attained between macroalgal biomass production and the paid environmental cost.

## 6.2. Economic feasibility

Biorefinery through concurrent production of valuable co-products that have wide industrial, medical, and nutritional applications simultaneously with biofuels has more significant potential towards circular economy. For example, by integrating seaweeds and fish farms, macroalgae can remove nitrogen, phosphorus and heavy metals from the pond [175,177] and oxygenate the water while utilizing the ammonia excreted by fish for algal growth [182]. In general, farming of seaweeds has additional socio-economic benefits, and is a vital industrial route as well as providing employment chances in developing countries. The social and economic dimensions of macroalgae farming were investigated by Valderrama et al. [93] who concluded that the net return to a family of four persons was higher than the international poverty line. However, due to the high labor demand and costly equipment required for cultivation of seaweeds, the economic value of the target product should be considered in order to be sufficient to make it worthwhile. For enhanced capacity of aquaculture to meet the economic feasibility, integration and expansion of macroalgae in marine aquaculture production has been proposed [183,184]. Despite the potential of numerous applications of macroalgal farms, it has not taken off globally due to the high production cost [185]. The manipulation costs, mainly coming from transportation and drying, should also be considered.

In order to reduce the transportation and drying costs, a mobile thermal decomposition unit can be built on-site with energy recycling (Fig. 5B). Heat recovery can be achieved via combustion of gases and biochar for biomass drying and thermal decomposition. In addition, the exothermal heat during pyrolysis can also be used to realize an energy self-balancing system in the large-scale plant. For such system, organic Rankine cycle will have a vital role as a simple, high reliable, and efficient route to convert low-grade waste heat to power [89,186]. In that context, Brigljević et al. [187] studied the economic feasibility of a large-scale poly-generation pyrolysis process of  $4 \times 10^5$  tons year<sup>-1</sup> of the dry brown seaweed *Saccharina japonica* as a feedstock to produce diesel-range hydrocarbons, power and heat with heat recycling through Rankine cycle. The sensitivity analysis parameters included internal economic parameters (fixed capital investment, internal rate of return, and income tax rate) and external economic parameters (the prices of seaweed, natural gas, power, acid, and hydrogen). The hydrogen price was considered as the selling price (Case 1) and the purchase price (Case 2). Comparison of four net present values (NPV) for both cases showed a similar increasing trend. However, Case 1 showed higher trend than Case 2, which was attributed to the higher capital investment in Case 1 (Fig. 6A). It was further confirmed by calculating the average return on investment (ROI) against NPV trends. At the same NPV value, results showed a notable higher ROI in Case 1 than Case 2 (Fig. 6B). Despite the low CO<sub>2</sub> emissions in the 2 studied cases, the study estimated higher CO<sub>2</sub> emissions in Case 1 than Case 2 (0.043 and 0.007 kg of CO<sub>2</sub> per kg of dry seaweed, respectively), which provide a beneficial environmental potential as CO<sub>2</sub> emissions are reduced by 7- to 45-fold compared to that produced from conventional crude oil (Fig. 6C). However, the estimated minimum product selling prices for Case 1 and Case 2 were US\$ 2.821 L<sup>-1</sup> and US\$ 2.847 L<sup>-1</sup>, respectively, which are higher than the current global average of diesel prices (US\$ 1.060 L<sup>-1</sup>) (Fig. 6C). Therefore, integration of seaweeds energy production with other industrial applications could enhance the process economy. For instance, seaweed aquaculture in combination with offshore wind energy production was suggested to reduce the costs of both offshore wind energy generation and seaweeds biomass production [188,189].

## 7. Future perspectives

Despite the positive environmental impacts and potential of seaweeds to produce crude bio-oil, the economic feasibility provided in the present study confirmed that the process is not economically feasible yet. The main challenge is to reduce the overall production cost and enhance the process economy. In addition, biofuel production from seaweeds encounters many technical challenges such as variation in seaweed growth and chemical composition depending on the season and geographical location [68,190]. Moreover, there is an urgent need to evaluate the impact of wild-harvesting on the marine ecosystems [191]. Therefore, developing advanced cultivation techniques is important in order to fulfil the specific characteristics of a certain algal species. Although some recent studies provided positive estimations on the cost of seaweed production for value-added products, much lower production cost must be achieved in case of biofuel. For that regard, combination of seaweed production, fish aquarium, phycoremediation, and offshore wind energy generation is expected to have synergetic interaction which turns the seaweed production into a profitable business. In addition, the economic feasibility of biofuel production from seaweeds can be further improved using an integrated biorefinery approach for simultaneous production of high-value products and biofuel, which enables the circular economy. Nowadays, the market for bioproducts from seaweeds is diverse, with a huge amount of residual biomass left behind. According to Tedesco and Daniels [191], 83–90% of the total seaweed-based industry are used as food, while the remaining 10–17% are used for bioproducts extraction, which produces huge amounts of biomass residues suitable for biofuel production.

Developing advanced conversion technologies is also important in

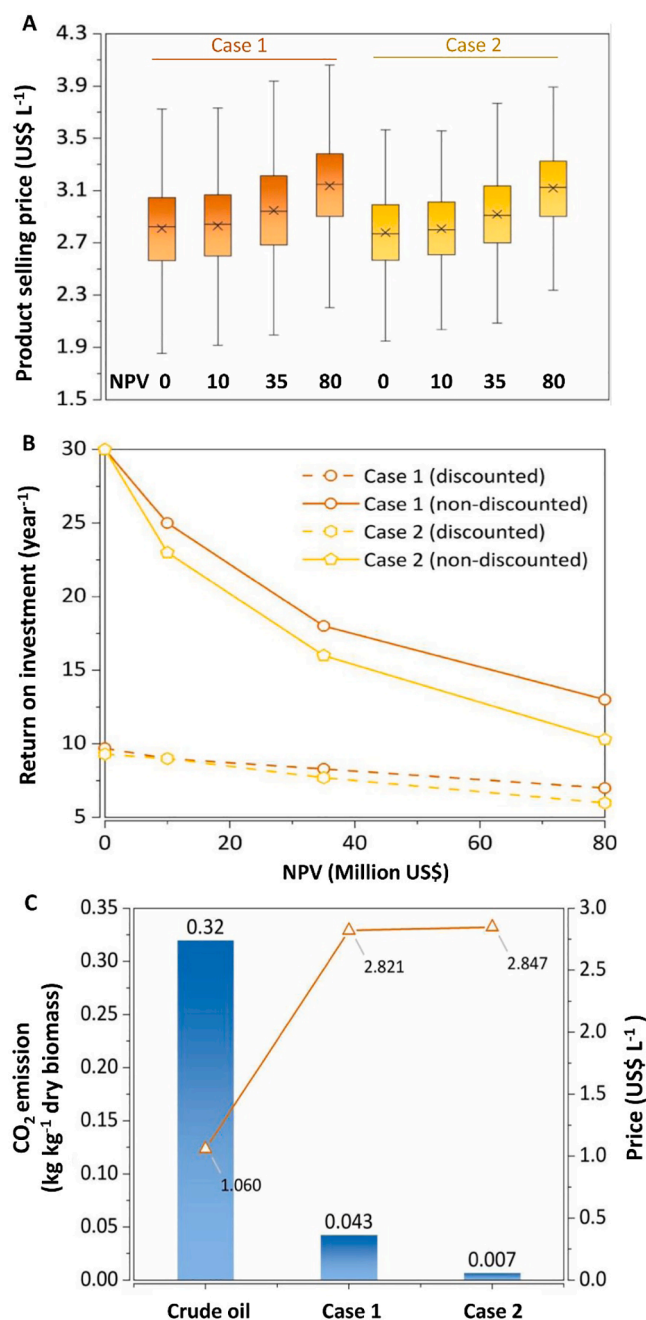


Fig. 6. A) Product selling price ranges at different net present values (NPV) of US\$ 0–80 mil (from left to right) at Case 1 and Case 2. B) Annual return on investment at different NPV for Case 1 and Case 2. C) CO<sub>2</sub> emissions (columns) and fuel price (line) of crude oil compared to biofuel produced from the two scenarios. Figures adopted from Ref. [187] after obtaining the required copyright permission from Elsevier.

order to enhance energy recovery from seaweeds. In that context, microwave pyrolysis was recently discussed as a promising technique for biomass conversion [162,192–194]. In addition, co-pyrolysis of seaweeds with high H and C feedstocks, such as waste plastic, could improve the yield and characteristics of the bio-oil [21,111,195]. Moreover, co-pyrolysis of seaweeds with cost-effective feedstocks such as fat, oil, and grease (FOG) could enhance the process economy. In general, developing the better economic models is essential to estimate how the cost of the integrated process varies with the characteristics of the final products, and the relative cost of the different crude bio-oil production routes versus algae-integrated technologies. Thus, more

studies should be conducted on the economic feasibility of pilot-scale conversion plants in order to provide a clear evaluation for the industrial applicability.

## 8. Conclusions

Crude bio-oil production from seaweeds could provide dual benefits of efficient biomass feedstock and save the environment from detrimental effects. Although many studies reported the efficiency of seaweeds for bio-oil production, large-scale production at low cost is the main challenge for that purpose. Combining algal farming, wastewater treatment, fish aquarium, wind power generation, synthesis of novel bioproducts and biofuel production is a promising strategy of bio-refinery towards circular economy that might reduce the overall cost. In addition, lower production cost can be achieved by developing new innovative technologies that reduce the cost at each processing step including; farm design, harvesting, pretreatment, transportation, feedstock selection, and conversion process. At the same time, environmental impacts of algal large-scale cultivation and/or extensive wild-harvesting need to be carefully considered. Overall, a balance must be attained between macroalgal bioenergy production, biofuel cost, and the environmental impacts.

## CRedit authorship contribution statement

**Shuang Wang:** Conceptualization, Methodology, Funding acquisition. **Shuang Zhao:** Methodology, Writing - original draft. **Benjamin Bernard Uzoejinwa:** Methodology, Writing - original draft. **Anqing Zheng:** Methodology, Writing - original draft. **Qingyuan Wang:** Writing - review & editing, Funding acquisition. **Jin Huang:** Writing - review & editing. **Abd El-Fatah Abomohra:** Conceptualization, Writing - review & editing, Funding acquisition.

## Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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## Appendix A. Supplementary data

Supplementary data to this article can be found online at <https://doi.org/10.1016/j.enconman.2020.113253>.

## References

- [1] Netravali AN, Chabba S. Composites get greener. *Mater Today* 2003;4:22–9.
- [2] Abomohra A-E-F, Jin W, Tu R, Han S-F, Eid M, Eladel H. Microalgal biomass production as a sustainable feedstock for biodiesel: current status and perspectives. *Renew Sustain Energy Rev* 2016;64:596–606.
- [3] Hoppe W, Bringezu S, Thonemann N. Comparison of global warming potential between conventionally produced and CO<sub>2</sub>-based natural gas used in transport versus chemical production. *J Clean Prod* 2016;121:231–7.
- [4] Bharathiraja B, Chakravarthy M, Kumar RR, Yogendran D, Yuvaraj D, Jayamuthunagai J, et al. Aquatic biomass (algae) as a future feed stock for bio-refineries: a review on cultivation, processing and products. *Renew Sustain Energy Rev* 2015;47:634–53.
- [5] Voelcker J. 1.2 billion vehicles on world's roads now, 2 billion by 2035: Report. *Green Car Reports* 2014;7.
- [6] Singh A, Olsen SI. A critical review of biochemical conversion, sustainability and life cycle assessment of algal biofuels. *Appl Energy* 2011;88:3548–55.

- [7] Elsayed M, Abomohra A-E-F, Ai P, Jin K, Fan Q, Zhang Y. Acetogenesis and methanogenesis liquid digestates for pretreatment of rice straw: a holistic approach for efficient biomethane production and nutrient recycling. *Energy Convers Manage* 2019;195:447–56.
- [8] Aresta M, Dibenedetto A, Barberio G. Utilization of macro-algae for enhanced CO<sub>2</sub> fixation and biofuels production: development of a computing software for an LCA study. *Fuel Process Technol* 2005;86:1679–93.
- [9] Shao W, Ebaid R, Abomohra AE, Shahan M. Enhancement of *Spirulina* biomass production and cadmium biosorption using combined static magnetic field. *Bioresour Technol* 2018. <https://doi.org/10.1016/j.biortech.2018.06.009>.
- [10] Abomohra A-E-F, El-Sheekh M, Hanelt D. Screening of marine microalgae isolated from the hypersaline Bardawil lagoon for biodiesel feedstock. *Renew Energy* 2017;101:1266–72.
- [11] Levine RB, Pinnarat T, Savage PE. Biodiesel production from wet algal biomass through in situ lipid hydrolysis and supercritical transesterification. *Energy Fuels* 2010;24:5235–43.
- [12] Amamou S, Sambusiti C, Monlau F, Dubreucq E, Barakat A. Mechano-enzymatic deconstruction with a new enzymatic cocktail to enhance enzymatic hydrolysis and bioethanol fermentation of two macroalgae species. *Molecules* 2018;23:174.
- [13] Gülzari ŞÖ, Lind V, Aasen IM, Steinshamn H. Effect of supplementing sheep diets with macroalgae species on in vivo nutrient digestibility, rumen fermentation and blood amino acid profile. *Animal* 2019;13:2792–801.
- [14] Kidanu WG, Trang PT, Yoon HH. Hydrogen and volatile fatty acids production from marine macroalgae by anaerobic fermentation. *Biotechnol Bioprocess Eng* 2017;22:612–9.
- [15] Elsayed M, Abomohra A-E-F, Ai P, Wang D, El-Mashad H, Zhang Y. Biorefining of rice straw by sequential fermentation and anaerobic digestion for bioethanol and/or biomethane production: comparison of structural properties and energy output. *Bioresour Technol* 2018;268:183–9.
- [16] Hagos K, Zong J, Li D, Liu C, Lu X. Anaerobic co-digestion process for biogas production: progress, challenges and perspectives. *Renew Sustain Energy Rev* 2017;76:1485–96.
- [17] Tabatabaei M, Aghbashlo M, Dehghani M, Panahi HKS, Mollahosseini A, Hosseini M, et al. Reactor technologies for biodiesel production and processing: a review. *Prog Energy Combust Sci* 2019;74:239–303.
- [18] Ha GS, El-Dalatony MM, Kim DH, Salama ES, Kurade MB, Roh HS, et al. Biocomponent-based microalgal transformations into biofuels during the pretreatment and fermentation process. *Bioresour Technol* 2020;302. <https://doi.org/10.1016/j.biortech.2020.122809>.
- [19] Abomohra AEF, El-Naggar AH, Baeshen AA. Potential of macroalgae for biodiesel production: Screening and evaluation studies. *J Biosci Bioeng* 2018;125:231–7.
- [20] Abomohra AE-F, Elsayed M, Esakkimuthu S, El-Sheekh M, Hanelt D. Potential of fat, oil and grease (FOG) for biodiesel production: a critical review on the recent progress and future perspectives. *Prog Energy Combust Sci* 2020;81. 100868.
- [21] Xu S, Cao B, Uzojinwa BB, Odey EA, Wang S, Shang H, et al. Synergistic effects of catalytic co-pyrolysis of macroalgae with waste plastics. *Process Saf Environ Prot* 2020;137:34–48.
- [22] Tabatabaei M, Aghbashlo M, Valijanian E, Kazemi Shariat Panahi H, Nizami AS, Ghanavati H, et al. A comprehensive review on recent biological innovations to improve biogas production, Part 1: upstream strategies. *Renew Energy* 2020;146:1204–20. <https://doi.org/10.1016/j.renene.2019.07.037>.
- [23] Fakayode OA, Aboagarib EAA, Zhou C, Ma H. Co-pyrolysis of lignocellulosic and macroalgae biomasses for the production of biochar—a review. *Bioresour Technol* 2020;297:122408.
- [24] Lee XJ, Ong HC, Gan YY, Chen W-H, Mahlia TMI. State of art review on conventional and advanced pyrolysis of macroalgae and microalgae for biochar, bio-oil and bio-syngas production. *Energy Convers Manage* 2020;210:112707.
- [25] Ma C, Geng J, Zhang D, Ning X. Non-catalytic and catalytic pyrolysis of *Ulva prolifera* macroalgae for production of quality bio-oil. *J Energy Inst* 2020;93:303–11.
- [26] Soltanian S, Aghbashlo M, Almasi F, Hosseinzadeh-Bandbafha H, Nizami A-S, Ok YS, et al. A critical review of the effects of pretreatment methods on the exergetic aspects of lignocellulosic biofuels. *Energy Convers Manage* 2020;212:112792.
- [27] Biswas B, Kumar A, Fernandes AC, Saini K, Negi S, Muraleedharan UD, et al. Solid base catalytic hydrothermal liquefaction of macroalgae: effects of process parameter on product yield and characterization. *Bioresour Technol* 2020;123232.
- [28] Ma C, Geng J, Zhang D, Ning X. Hydrothermal liquefaction of macroalgae: influence of zeolites based catalyst on products. *J Energy Inst* 2020;93:581–90.
- [29] Parsa M, Nourani M, Baghdadi M, Hosseinzadeh M, Pejman M. Biochars derived from marine macroalgae as a mesoporous by-product of hydrothermal liquefaction process: characterization and application in wastewater treatment. *J Water Process Eng* 2019;32:100942.
- [30] Yan L, Wang Y, Li J, Zhang Y, Ma L, Fu F, et al. Hydrothermal liquefaction of *Ulva prolifera* macroalgae and the influence of base catalysts on products. *Bioresour Technol* 2019;292:121286.
- [31] Cao J, Ma Y. Pyrolysis and gasification of macroalgae *Enteromorpha prolifera* under a CO<sub>2</sub> atmosphere using the thermogravimetry–Fourier transform infrared spectroscopy technique. *Prog React Kinet Mech* 2019;44:132–42.
- [32] He Z, Saw WL, Lane DJ, van Eyk PJ, de Nys R, Nathan GJ, et al. The ash-quartz sand interaction behaviours during steam gasification or combustion of a freshwater and a marine species of macroalgae. *Fuel* 2020;263:116621.
- [33] Lane DJ, Truong E, Larizza F, Chiew P, de Nys R, van Eyk PJ. Effect of hydrothermal carbonization on the combustion and gasification behavior of agricultural residues and macroalgae: devolatilization characteristics and char reactivity. *Energy Fuels* 2017;32:4149–59.
- [34] Aitken D, Bulboa C, Godoy-Faundez A, Turrión-Gómez JL, Antizar-Ladislao B. Life cycle assessment of macroalgae cultivation and processing for biofuel production. *J Clean Prod* 2014;75:45–56.
- [35] Tan IS, Lee KT. Solid acid catalysts pretreatment and enzymatic hydrolysis of macroalgae cellulosic residue for the production of bioethanol. *Carbohydr Polym* 2015;124:311–21.
- [36] Jiang R, Ingle KN, Golberg A. Macroalgae (seaweed) for liquid transportation biofuel production: what is next? *Algal Res* 2016;14:48–57.
- [37] Kraan S. Mass-cultivation of carbohydrate rich macroalgae, a possible solution for sustainable biofuel production. *Mitig Adapt Strateg Glob Change* 2013;18:27–46.
- [38] Jung KA, Lim S-R, Kim Y, Park JM. Potentials of macroalgae as feedstocks for biorefinery. *Bioresour Technol* 2013;135:182–90.
- [39] Bucholc K, Szymczak-Żyła M, Lubecki L, Zamojska A, Hapter P, Tjernström E, et al. Nutrient content in macrophyta collected from southern Baltic Sea beaches in relation to eutrophication and biogas production. *Sci Total Environ* 2014;473:298–307.
- [40] Roesijadi G, Jones SB, Snowden-Swan LJ, Zhu Y. Macroalgae as a biomass feedstock: a preliminary analysis. Pacific Northwest National Lab.(PNNL), Richland, WA (United States); 2010.
- [41] FAO. Fisheries and aquaculture information and statistics services 2014. <http://www.fao.org/figis/> (accessed March 26, 2019).
- [42] Rebours C, Marinho-Soriano E, Zertuche-González JA, Hayashi L, Vásquez JA, Kradolfer P, et al. Seaweeds: an opportunity for wealth and sustainable livelihood for coastal communities. *J Appl Phycol* 2014;26:1939–51.
- [43] Ventura MR, Castañón JIR. The nutritive value of seaweed (*Ulva lactuca*) for goats. *Small Rumin Res* 1998;29:325–7.
- [44] Ghadiryanfar M, Rosentrater KA, Keyhani A, Omid M. A review of macroalgae production, with potential applications in biofuels and bioenergy. *Renew Sustain Energy Rev* 2016;54:473–81.
- [45] Verma P, Kumar M, Mishra G, Sahoo D. Multivariate analysis of fatty acid and biochemical constituents of seaweeds to characterize their potential as bioresource for biofuel and fine chemicals. *Bioresour Technol* 2017;226:132–44.
- [46] Yuan C, Wang S, Cao B, Hu Y, Abomohra AEF, Wang Q, et al. Optimization of hydrothermal co-liquefaction of seaweeds with lignocellulosic biomass: merging 2nd and 3rd generation feedstocks for enhanced bio-oil production. *Energy* 2019;173:413–22.
- [47] Cao B, Sun Y, Guo J, Wang S, Yuan J, Esakkimuthu S, et al. Synergistic effects of co-pyrolysis of macroalgae and polyvinyl chloride on bio-oil/bio-char properties and transferring regularity of chlorine. *Fuel* 2019;246:319–29. <https://doi.org/10.1016/j.fuel.2019.02.037>.
- [48] Aravind S, Kumar PS, Kumar NS, Siddarth N. Conversion of green algal biomass into bioenergy by pyrolysis. A review. *Environ Chem Lett* 2020;1–21.
- [49] Xu S, Wang S, Zhang Z, Li C, Jiang X. Study on pore structure of seaweed particles after combustion. *J Energy Resour Technol* 2016;138.
- [50] Hu Y, Wang S, Wang Q, He Z, Lin X, Xu S, et al. Effect of different pretreatments on the thermal degradation of seaweed biomass. *Proc Combust Inst* 2017;36:2271–81.
- [51] Sudhakar MP, Kumar BR, Mathimani T, Arunkumar K. A review on bioenergy and bioactive compounds from microalgae and macroalgae-sustainable energy perspective. *J Clean Prod* 2019;228:1320–33.
- [52] Panahi HKS, Dehghani M, Aghbashlo M, Karimi K, Tabatabaei M. Shifting fuel feedstock from oil wells to sea: Iran outlook and potential for biofuel production from brown macroalgae (ochrophyta; phaeophyceae). *Renew Sustain Energy Rev* 2019;112:626–42.
- [53] Zainan NH, Srivatsa SC, Li F, Bhattacharya S. Quality of bio-oil from catalytic pyrolysis of microalgae *Chlorella vulgaris*. *Fuel* 2018;223:12–9.
- [54] Panahi HKS, Tabatabaei M, Aghbashlo M, Dehghani M, Rehan M, Nizami A-S. Recent updates on the production and upgrading of bio-crude oil from microalgae. *Bioresour Technol Reports* 2019;7:100216.
- [55] Jena U, Das KC. Comparative evaluation of thermochemical liquefaction and pyrolysis for bio-oil production from microalgae. *Energy Fuels* 2011;25:5472–82.
- [56] Priharto N, Ronsse F, Prins W, Carleer R, Heeres HJ. Experimental studies on a two-step fast pyrolysis-catalytic hydrotreatment process for hydrocarbons from microalgae (*Nannochloropsis gaditana* and *Scenedesmus almeriensis*). *Fuel Process Technol* 2020;206:106466.
- [57] Andrade LA, Barbosa JM, Barrozo MAS, Vieira LGM. A comparative study of the behavior of *Chlamydomonas reinhardtii* and *Spirulina platensis* in solar catalytic pyrolysis. *Int J Energy Res* 2020;44:5397–411.
- [58] Anand V, Gautam R, Vinu R. Non-catalytic and catalytic fast pyrolysis of *Schizochytrium limacinum* microalgae. *Fuel* 2017;205:1–10.
- [59] Wei N, Quarterman J, Jin Y-S. Marine macroalgae: an untapped resource for producing fuels and chemicals. *Trends Biotechnol* 2013;31:70–7.
- [60] Alexandropoulou M, Antonopoulou G, Fragkou E, Ntaikou I, Lyberatos G. Fungal pretreatment of willow sawdust and its combination with alkaline treatment for enhancing biogas production. *J Environ Manage* 2017;203:704–13.
- [61] Khan MY, Dahot MU. Effect of various agriculture wastes and pure sugars on the production of single cell protein by *Penicillium expansum*. *World Appl Sci J* 2010;8:80–4.
- [62] Adams JM, Gallagher JA, Donnison IS. Fermentation study on *Saccharina latissima* for bioethanol production considering variable pre-treatments. *J Appl Phycol* 2009;21:569.
- [63] Horn SJ, Aasen IM, Østgaard K. Production of ethanol from mannitol by *Zymobacter palmarum*. *J Ind Microbiol Biotechnol* 2000;24:51–7.
- [64] Guiry MD. The seaweed site: Information on marine algae 2014.

- [65] Samaraweera AM, Vidanarachchi JK, Kurukulasuriya MS. Industrial applications of macroalgae. *Handb Mar Macroalgae* 2012;500–21.
- [66] Murdock JN, Wetzel DL. FT-IR microspectroscopy enhances biological and ecological analysis of algae. *Appl Spectrosc Rev* 2009;44:335–61.
- [67] Briand X, Morand P. Anaerobic digestion of *Ulva* sp. 1. Relationship between *Ulva* composition and methanisation. *J Appl Phycol* 1997;9:511.
- [68] Osman MEH, Abo-Shady AM, Elshobary ME, Abd El-Ghafar MO, Abomohra AE-F. Screening of seaweeds for sustainable biofuel recovery through sequential biodiesel and bioethanol production 2020. <https://doi.org/10.1007/s11356-020-09534-1>.
- [69] Devi IRP, Gowri VS. Biological treatment of aquaculture discharge waters by seaweeds. *J Control Pollut* 2007;23:135–40.
- [70] Aderhold D, Williams CJ, Edyvean RGJ. The removal of heavy-metal ions by seaweeds and their derivatives. *Bioresour Technol* 1996;58:1–6.
- [71] Xiong J-Q, Kurade MB, Jeon B-H. Can microalgae remove pharmaceutical contaminants from water? *Trends Biotechnol* 2018;36:30–44.
- [72] Gou N, Yuan S, Lan J, Gao C, Alshawabkeh AN, Gu AZ. A quantitative toxicogenomics assay reveals the evolution and nature of toxicity during the transformation of environmental pollutants. *Environ Sci Technol* 2014;48:8855–63.
- [73] Murugesan K, Chang Y-Y, Kim Y-M, Jeon J-R, Kim E-J, Chang Y-S. Enhanced transformation of triclosan by laccase in the presence of redox mediators. *Water Res* 2010;44:298–308.
- [74] Zietzschmann F, Stützer C, Jekel M. Granular activated carbon adsorption of organic micro-pollutants in drinking water and treated wastewater—aligning breakthrough curves and capacities. *Water Res* 2016;92:180–7.
- [75] Afreen S, Shamsi TN, Baig MA, Ahmad N, Fatima S, Qureshi MI, et al. A novel multicopper oxidase (laccase) from cyanobacteria: purification, characterization with potential in the decolorization of anthraquinonic dye. *PLoS ONE* 2017;12:e0175144.
- [76] Latinwo GK, Jimoda LA, Agarry SE, Adeniran JA. Biosorption of some heavy metals from Textile Wastewater by Green Seaweed Biomass. *Univers J Environ Res Technol* 2015;5.
- [77] Ungureanu G, Santos SCR, Volf I, Boaventura RAR, Botelho CMS. Biosorption of antimony oxyanions by brown seaweeds: batch and column studies. *J Environ Chem Eng* 2017;5:3463–71.
- [78] Navarro AE, Portales RF, Sun-Kou MR, Llanos BP. Effect of pH on phenol biosorption by marine seaweeds. *J Hazard Mater* 2008;156:405–11.
- [79] Flores-Chaparro CE, Ruiz LFC, de la Torre MCA, Huerta-Diaz MA, Rangel-Mendez JR. Biosorption removal of benzene and toluene by three dried macroalgae at different ionic strength and temperatures: algae biochemical composition and kinetics. *J Environ Manage* 2017;193:126–35.
- [80] Bai X, Acharya K. Algae-mediated removal of selected pharmaceutical and personal care products (PPCPs) from Lake Mead water. *Sci Total Environ* 2017;581:734–40.
- [81] Matamoros V, Uggetti E, García J, Bayona JM. Assessment of the mechanisms involved in the removal of emerging contaminants by microalgae from wastewater: a laboratory scale study. *J Hazard Mater* 2016;301:197–205.
- [82] Hom-Diaz A, Jaén-Gil A, Bello-Laserna I, Rodríguez-Mozaz S, Vicent T, Barceló D, et al. Performance of a microalgal photobioreactor treating toilet wastewater: pharmaceutically active compound removal and biomass harvesting. *Sci Total Environ* 2017;592:1–11.
- [83] Ding T, Yang M, Zhang J, Yang B, Lin K, Li J, et al. Toxicity, degradation and metabolic fate of ibuprofen on freshwater diatom *Navicula* sp. *J Hazard Mater* 2017;330:127–34.
- [84] Ceylan S, Goldfarb JL. Green tide to green fuels: TG-FTIR analysis and kinetic study of *Ulva* prolifera pyrolysis. *Energy Convers Manage* 2015;101:263–70.
- [85] Kan T, Grierson S, De Nys R, Strezov V. Comparative assessment of the thermochemical conversion of freshwater and marine micro-and macroalgae. *Energy Fuels* 2014;28:104–14.
- [86] Lin KC, Lin Y-C, Hsiao Y-H. Microwave plasma studies of *Spirulina* algae pyrolysis with relevance to hydrogen production. *Energy* 2014;64:567–74.
- [87] Demirbaş A. Biomass resource facilities and biomass conversion processing for fuels and chemicals. *Energy Convers Manage* 2001;42:1357–78.
- [88] Demirbaş A. Analysis of liquid products from biomass via flash pyrolysis. *Energy Sources* 2002;24:337–45.
- [89] Wang S, Yerkebulan M, Abomohra AEF, El-Khodary S, Wang Q. Microalgae harvest influences the energy recovery: a case study on chemical flocculation of *Scenedesmus obliquus* for biodiesel and crude bio-oil production. *Bioresour Technol* 2019;286:121371.
- [90] Yang J, Rizkiana J, Widayatno WB, Karnjanakom S, Kaewpanha M, Hao X, et al. Fast co-pyrolysis of low density polyethylene and biomass residue for oil production. *Energy Convers Manage* 2016;120:422–9.
- [91] Burton T, Lyons H, Lerat Y, Stanley M, Rasmussen MB. A review of the potential of marine algae as a source of biofuel in Ireland 2009.
- [92] Ai P, Zhang X, Ran Y, Meng L, Elsayed M, Fan Q, et al. Biomass briquetting reduces the energy loss during long-term ensiling and enhances anaerobic digestion: a case study on rice straw. *Bioresour Technol* 2019;292:121912.
- [93] Valderrama D, Cai J, Hishamunda N, Ridler N. Social and economic dimensions of carrageenan seaweed farming 2013.
- [94] Lightfoot DG, Raghavan GSV. Combined fields dewatering of seaweed (*Nereocystis luetkeana*). *Trans ASAE* 1994;37:899–906.
- [95] Oswald WJ. Large-scale algal culture systems (engineering aspects). *Micro-Algal Biotechnol* 1988:357–94.
- [96] Ross AB, Jones JM, Kubacki ML, Bridgeman T. Classification of macroalgae as fuel and its thermochemical behaviour. *Bioresour Technol* 2008;99:6494–504.
- [97] Ross AB, Anastasakis K, Kubacki M, Jones JM. Investigation of the pyrolysis behaviour of brown algae before and after pre-treatment using PY-GC/MS and TGA. *J Anal Appl Pyrolysis* 2009;85:3–10.
- [98] Cao B, Wang S, Hu Y, Abomohra AEF, Qian L, He Z, et al. Effect of washing with diluted acids on *Enteromorpha clathrata* pyrolysis products: towards enhanced bio-oil from seaweeds. *Renew Energy* 2019;138:29–38. <https://doi.org/10.1016/j.renene.2019.01.084>.
- [99] Eom I-Y, Kim K-H, Kim J-Y, Lee S-M, Yeo H-M, Choi I-G, et al. Characterization of primary thermal degradation features of lignocellulosic biomass after removal of inorganic metals by diverse solvents. *Bioresour Technol* 2011;102:3437–44.
- [100] Zhang S, Dong Q, Zhang L, Xiong Y, Liu X, Zhu S. Effects of water washing and torrefaction pretreatments on rice husk pyrolysis by microwave heating. *Bioresour Technol* 2015;193:442–8.
- [101] Ly HV, Kim S-S, Kim J, Choi JH, Woo HC. Effect of acid washing on pyrolysis of *Cladophora socialis* alga in microtubing reactor. *Energy Convers Manage* 2015;106:260–7.
- [102] Choi J, Choi J-W, Suh DJ, Ha J-M, Hwang JW, Jung HW, et al. Production of brown algae pyrolysis oils for liquid biofuels depending on the chemical pretreatment methods. *Energy Convers Manage* 2014;86:371–8.
- [103] Chee S-Y, Wong P-K, Wong C-L. Extraction and characterisation of alginate from brown seaweeds (Fucales, Phaeophyceae) collected from Port Dickson, Peninsular Malaysia. *J Appl Phycol* 2011;23:191–6.
- [104] Rodríguez-Jasso RM, Mussatto SI, Pastrana L, Aguilar CN, Teixeira JA. Extraction of sulfated polysaccharides by autohydrolysis of brown seaweed *Fucus vesiculosus*. *J Appl Phycol* 2013;25:31–9.
- [105] Bae YJ, Ryu C, Jeon J-K, Park J, Suh DJ, Suh Y-W, et al. The characteristics of bio-oil produced from the pyrolysis of three marine macroalgae. *Bioresour Technol* 2011;102:3512–20.
- [106] Ale MT, Mikkelsen JD, Meyer AS. Designed optimization of a single-step extraction of fucose-containing sulfated polysaccharides from *Sargassum* sp. *J Appl Phycol* 2012;24:715–23.
- [107] Park K, Cho E, In M-J, Kim DC, Chae HJ. Physicochemical properties and bioactivity of brown seaweed fucoidan prepared by ultra high pressure-assisted enzyme treatment. *Korean J Chem Eng* 2012;29:221–7.
- [108] Wang S, Uzojejinwa BB, Abomohra A-E-F, Wang Q, He Z, Feng Y, et al. Characterization and pyrolysis behavior of the green microalga *Microactinium conductrix* grown in lab-scale tubular photobioreactor using Py-GC/MS and TGA/MS. *J Anal Appl Pyrolysis* 2018;135:340–9.
- [109] Cha JS, Park SH, Jung S-C, Ryu C, Jeon J-K, Shin M-C, et al. Production and utilization of biochar: a review. *J Ind Eng Chem* 2016;40:1–15.
- [110] Ali I, Bahadar A. Red Sea seaweed (*Sargassum* spp.) pyrolysis and its devolatilization kinetics. *Algal Res* 2017;1:89–97.
- [111] Uzojejinwa BB, He X, Wang S, Abomohra A-E-F, Hu Y, Wang Q. Co-pyrolysis of biomass and waste plastics as a thermochemical conversion technology for high-grade biofuel production: Recent progress and future directions elsewhere worldwide. *Energy Convers Manage* 2018;163:468–92.
- [112] Balat M, Balat M, Kirtay E, Balat H. Main routes for the thermo-conversion of biomass into fuels and chemicals. Part 1: pyrolysis systems. *Energy Convers Manage* 2009;50:3147–57.
- [113] Astrup T, Billitewski B. Pyrolysis and gasification. *Solid Waste Technol. Manag., Wiley*; 2011, pp. 502–12.
- [114] Koshtikanawuth K, Sattler ML, Dennis B. Pyrolysis of macroalgae and polystyrene: a review. *Curr Sustain Energy Reports* 2014;1:121–8.
- [115] Moreno-Piraján JC, Gómez-Cruz R, García-Cuello VS, Giraldo L. Binary system Cu (II)/Pb (II) adsorption on activated carbon obtained by pyrolysis of cow bone study. *J Anal Appl Pyrolysis* 2010;89:122–8.
- [116] Mullen CA, Boateng AA, Goldberg NM, Lima IM, Laird DA, Hicks KB. Bio-oil and bio-char production from corn cobs and stover by fast pyrolysis. *Biomass Bioenergy* 2010;34:67–74.
- [117] Ibrahim MM, Tong C, Hu K, Zhou B, Xing S, Mao Y. Biochar-fertilizer interaction modifies N-sorption, enzyme activities and microbial functional abundance regulating nitrogen retention in rhizosphere soil. *Sci Total Environ* 2020;140065.
- [118] Gan YY, Ong HC, Show PL, Ling TC, Chen W-H, Yu KL, et al. Torrefaction of microalgal biochar as potential coal fuel and application as bio-adsorbent. *Energy Convers Manage* 2018;165:152–62.
- [119] Zhu X, Luo Z, Diao R, Zhu X. Combining torrefaction pretreatment and co-pyrolysis to upgrade biochar derived from bio-oil distillation residue and walnut shell. *Energy Convers Manage* 2019;199:111970.
- [120] Zhang L, Lim EY, Loh K-C, Ok YS, Lee JTE, Shen Y, et al. Biochar enhanced thermophilic anaerobic digestion of food waste: focusing on biochar particle size, microbial community analysis and pilot-scale application. *Energy Convers Manage* 2020;209:112654.
- [121] Pedrazzi S, Santunione G, Minarelli A, Allesina G. Energy and biochar co-production from municipal green waste gasification: a model applied to a landfill in the north of Italy. *Energy Convers Manage* 2019;187:274–82.
- [122] Park HJ, Dong J-I, Jeon J-K, Park Y-K, Yoo K-S, Kim S-S, et al. Effects of the operating parameters on the production of bio-oil in the fast pyrolysis of Japanese larch. *Chem Eng J* 2008;143:124–32.
- [123] Jung S-H, Kang B-S, Kim J-S. Production of bio-oil from rice straw and bamboo sawdust under various reaction conditions in a fast pyrolysis plant equipped with a fluidized bed and a char separation system. *J Anal Appl Pyrolysis* 2008;82:240–7.
- [124] Wang S, Wang Q, Jiang X, Han X, Ji H. Compositional analysis of bio-oil derived from pyrolysis of seaweed. *Energy Convers Manage* 2013;68:273–80.

- [125] Choi JH, Kim S-S, Suh DJ, Jang E-J, Min K-I, Woo HC. Characterization of the bio-oil and bio-char produced by fixed bed pyrolysis of the brown alga *Saccharina japonica*. Korean J Chem Eng 2016;33:2691–8.
- [126] Trinh TN, Jensen PA, Dam-Johansen K, Knudsen NO, Sørensen HR, Hvilsted S. Comparison of lignin, macroalgae, wood, and straw fast pyrolysis. Energy Fuels 2013;27:1399–409.
- [127] Ferrera-Lorenzo N, Fuente E, Suárez-Ruiz I, Gil RR, Ruiz B. Pyrolysis characteristics of a macroalgae solid waste generated by the industrial production of Agar-Agar. J Anal Appl Pyrolysis 2014;105:209–16.
- [128] Norouzi O, Jafarian S, Safari F, Tavasoli A, Nejati B. Promotion of hydrogen-rich gas and phenolic-rich bio-oil production from green macroalgae *Cladophora glomerata* via pyrolysis over its bio-char. Bioresour Technol 2016;219:643–51.
- [129] Chiodo V, Zafarana G, Maisano S, Freni S, Urbani F. Pyrolysis of different biomass: direct comparison among *Posidonia Oceanica*, *Lacustrine Alga* and *White-Pine*. Fuel 2016;164:220–7.
- [130] Yanik J, Stahl R, Troeger N, Sinag A. Pyrolysis of algal biomass. J Anal Appl Pyrolysis 2013;103:134–41.
- [131] Choi J-W, Choi JH, Suh DJ, Kim H. Feasibility of *Laminaria japonica* as a feedstock for fast pyrolysis in a bubbling fluidized-bed reactor. J Anal Appl Pyrolysis 2015;112:141–9.
- [132] Attri P, Yusupov M, Park JH, Lingamdinne LP, Koduru JR, Shiratani M, et al. Mechanism and comparison of needle-type non-thermal direct and indirect atmospheric pressure plasma jets on the degradation of dyes. Sci Rep 2016;6: 34419.
- [133] Jiang D, Xia Z, Wang S, Li H, Gong X, Yuan C, et al. Mechanism research on catalytic pyrolysis of sulfated polysaccharide using ZSM-5 catalysts by Py-GC/MS and density functional theory studies. J Anal Appl Pyrolysis 2019;143. <https://doi.org/10.1016/j.jaap.2019.104680>.
- [134] Eschenbacher A, Jensen PA, Henriksen UB, Ahrenfeldt J, Jensen CD, Li C, et al. Catalytic upgrading of tars generated in a 100 kWth low temperature circulating fluidized bed gasifier for production of liquid bio-fuels in a polygeneration scheme. Energy Convers Manage 2020;207:112538.
- [135] Plis A, Lasek J, Skawińska A, Zuwała J. Thermochemical and kinetic analysis of the pyrolysis process in *Cladophora glomerata* alga. J Anal Appl Pyrolysis 2015; 115:166–74.
- [136] Wang S, Cao B, Abomohra A-E-F, Hu Y, Wang Q, He Z, et al. Comparative Study of Combustion Properties of Two Seaweeds in a Batch Fluidized Bed. Combust Sci Technol 2018;190:755–69. <https://doi.org/10.1080/00102202.2017.1407761>.
- [137] Li D, Chen L, Yi X, Zhang X, Ye N. Pyrolytic characteristics and kinetics of two brown algae and sodium alginate. Bioresour Technol 2010;101:7131–6.
- [138] Kebelmann K, Hornung A, Karsten U, Griffiths G. Thermo-chemical behaviour and chemical product formation from Polar seaweeds during intermediate pyrolysis. J Anal Appl Pyrolysis 2013;104:131–8.
- [139] Uzojejinwa BB, He X, Wang S, Abomohra A-E-F, Hu Y, He Z, et al. Co-pyrolysis of macroalgae and lignocellulosic biomass. J Therm Anal Calorim 2019;136: 2001–16.
- [140] Wang S, Cao B, Liu X, Xu L, Hu Y, Afonaa-Mensah S, et al. A comparative study on the quality of bio-oil derived from green macroalgae *Enteromorpha clathrata* over metal modified ZSM-5 catalysts. Bioresour Technol 2018;256:446–55. <https://doi.org/10.1016/j.biortech.2018.01.134>.
- [141] Wang S, Shang H, Abomohra AE-F, Wang Q. One-step conversion of microalgae to alcohols and esters through co-pyrolysis with biodiesel-derived glycerol. Energy Convers Manage 2019;198:111792.
- [142] Wang S, Jiang D, Cao B, Hu Y, Yuan C, Wang Q, et al. Study on the interaction effect of seaweed bio-coke and rice husk volatiles during co-pyrolysis. J Anal Appl Pyrolysis 2018;132:111–22. <https://doi.org/10.1016/j.jaap.2018.03.009>.
- [143] Wang S, Jiang XM, Wang N, Yu LJ, Li Z, He PM. Research on pyrolysis characteristics of seaweed. Energy Fuels 2007;21:3723–9.
- [144] Van Heemst JDH, Peulve S, De Leeuw JW. Novel algal polyphenolic biomacromolecules as significant contributors to resistant fractions of marine dissolved and particulate organic matter. Org Geochem 1996;24:629–40.
- [145] Sajdak M, Muzyka R, Hrabak J, Stowik K. Use of plastic waste as a fuel in the co-pyrolysis of biomass: part III: optimisation of the co-pyrolysis process. J Anal Appl Pyrolysis 2015;112:298–305.
- [146] Xu S, Elsayed M, Ismail GA, Li C, Wang S, Abomohra AEF. Evaluation of bioethanol and biodiesel production from *Scenedesmus obliquus* grown in biodiesel waste glycerol: A sequential integrated route for enhanced energy recovery. Energy Convers Manage 2019;197. <https://doi.org/10.1016/j.enconman.2019.111907>.
- [147] Xu S, Uzojejinwa BB, Wang S, Hu Y, Qian L, Liu L, et al. Study on co-pyrolysis synergistic mechanism of seaweed and rice husk by investigation of the characteristics of char/coke. Renew Energy 2019;132:527–42. <https://doi.org/10.1016/j.renene.2018.08.025>.
- [148] Kim Y-M, Lee HW, Choi SJ, Jeon J-K, Park SH, Jung S-C, et al. Catalytic co-pyrolysis of polypropylene and *Laminaria japonica* over zeolitic materials. Int J Hydrogen Energy 2017;42:18434–41.
- [149] Wang S, Xia Z, Hu Y, He Z, Uzojejinwa BB, Wang Q, et al. Co-pyrolysis mechanism of seaweed polysaccharides and cellulose based on macroscopic experiments and molecular simulations. Bioresour Technol 2017;228:305–14.
- [150] Cao B, Xia Z, Wang S, Abomohra A-E-F, Cai N, Hu Y, et al. A study on catalytic co-pyrolysis of cellulose with seaweeds polysaccharides over ZSM-5: Towards high-quality biofuel production. J Anal Appl Pyrolysis 2018. <https://doi.org/10.1016/j.jaap.2018.07.020>.
- [151] Lee HW, Choi SJ, Park SH, Jeon J-K, Jung S-C, Joo SH, et al. Catalytic conversion of *Laminaria japonica* over microporous zeolites. Energy 2014;66:2–6.
- [152] Lorenzetti C, Conti R, Fabbri D, Yanik J. A comparative study on the catalytic effect of H-ZSM5 on upgrading of pyrolysis vapors derived from lignocellulosic and proteinaceous biomass. Fuel 2016;166:446–52.
- [153] Lee HW, Jeon J-K, Park SH, Jeong K-E, Chae H-J, Park Y-K. Catalytic pyrolysis of *Laminaria japonica* over nanoporous catalysts using Py-GC/MS. Nanoscale Res Lett 2011;6:500.
- [154] Mihalek DJ, Mullen CA, Boateng AA. Screening acidic zeolites for catalytic fast pyrolysis of biomass and its components. J Anal Appl Pyrolysis 2011;92:224–32.
- [155] Kim B-S, Jeong CS, Kim JM, Bin PS, Park SH, Jeon J-K, et al. Ex situ catalytic upgrading of lignocellulosic biomass components over vanadium contained H-MCM-41 catalysts. Catal Today 2016;265:184–91.
- [156] Zhang B, Zhong Z, Ding K, Song Z. Production of aromatic hydrocarbons from catalytic co-pyrolysis of biomass and high density polyethylene: analytical Py-GC/MS study. Fuel 2015;139:622–8.
- [157] Kim J-Y, Lee JH, Park J, Kim JK, An D, Song IK, et al. Catalytic pyrolysis of lignin over HZSM-5 catalysts: effect of various parameters on the production of aromatic hydrocarbon. J Anal Appl Pyrolysis 2015;114:273–80.
- [158] Casoni AI, Nievas ML, Moyano EL, Álvarez M, Diez A, Dennehy M, et al. Catalytic pyrolysis of cellulose using MCM-41 type catalysts. Appl Catal A Gen 2016;514: 235–40.
- [159] Jeon M-J, Jeon J-K, Suh DJ, Park SH, Sa YJ, Joo SH, et al. Catalytic pyrolysis of biomass components over mesoporous catalysts using Py-GC/MS. Catal Today 2013;204:170–8.
- [160] Hu Y, Wang H, Lakshmikanand M, Wang S, Wang Q, He Z, et al. Catalytic co-pyrolysis of seaweeds and cellulose using mixed ZSM-5 and MCM-41 for enhanced crude bio-oil production. J Therm Anal Calorim 2020. <https://doi.org/10.1007/s10973-020-09291-w>.
- [161] Dai M, Yu Z, Fang S, Ma X. Behaviors, product characteristics and kinetics of catalytic co-pyrolysis spirulina and oil shale. Energy Convers Manage 2019;192: 1–10. <https://doi.org/10.1016/j.enconman.2019.04.032>.
- [162] Lam SS, Mahari WAW, Ok YS, Peng W, Chong CT, Ma NL, et al. Microwave vacuum pyrolysis of waste plastic and used cooking oil for simultaneous waste reduction and sustainable energy conversion: recovery of cleaner liquid fuel and techno-economic analysis. Renew Sustain Energy Rev 2019;115:109359.
- [163] Tang Z, Chen W, Chen Y, Yang H, Chen H. Co-pyrolysis of microalgae and plastic: Characteristics and interaction effects. Bioresour Technol 2019;274:145–52.
- [164] Zhang X, Lei H, Chen S, Wu J. Catalytic co-pyrolysis of lignocellulosic biomass with polymers: a critical review. Green Chem 2016;18:4145–69.
- [165] Brebu M, Yanik J, Uysal T, Vasile C. Thermal and catalytic degradation of grape seeds/polyethylene waste mixture. Cellul Chem Technol 2014;48:665–74.
- [166] Hu G, Li J, Zhang X, Li Y. Investigation of waste biomass co-pyrolysis with petroleum sludge using a response surface methodology. J Environ Manage 2017; 192:234–42.
- [167] Siddiqui MTH, Nizamuddin S, Mubarak NM, Shirin K, Aijaz M, Hussain M, et al. Characterization and process optimization of biochar produced using novel biomass, waste pomegranate peel: a response surface methodology approach. Waste Biomass Valorization 2019;10:521–32.
- [168] Rojas-Mayorga CK, Bonilla-Petriciolet A, Aguayo-Villarreal IA, Hernandez-Montoya V, Moreno-Virgen MR, Tovar-Gómez R, et al. Optimization of pyrolysis conditions and adsorption properties of bone char for fluoride removal from water. J Anal Appl Pyrolysis 2013;104:10–8.
- [169] Ren X, Gou J, Wang W, Li Q, Chang J, Li B. Optimization of bark fast pyrolysis for the production of phenol-rich bio-oil. BioResources 2013;8:6481–92.
- [170] Dufour J, Iribarren D. Life cycle assessment of biodiesel production from free fatty acid-rich wastes. Renew Energy 2012;38:155–62.
- [171] Elsayed M, Ran Y, Ai P, Azab M, Mansour A, Jin K, et al. Innovative integrated approach of biofuel production from agricultural wastes by anaerobic digestion and black soldier fly larvae. J Clean Prod 2020. 121495.
- [172] Gao K, McKinley KR. Use of macroalgae for marine biomass production and CO<sub>2</sub> remediation: a review. J Appl Phycol 1994;6:45–60.
- [173] Dave A, Huang Y, Rezvani S, McIlveen-Wright D, Novaes M, Hewitt N. Techno-economic assessment of biofuel development by anaerobic digestion of European marine cold-water seaweeds. Bioresour Technol 2013;135:120–7.
- [174] Tabassum MR, Xia A, Murphy JD. Biomethane production from various segments of brown seaweed. Energy Convers Manage 2018;174:855–62.
- [175] Alvarado-Morales M, Boldrin A, Karakashev DB, Holdt SL, Angelidaki I, Astrup T. Life cycle assessment of biofuel production from brown seaweed in Nordic conditions. Bioresour Technol 2013;129:92–9.
- [176] Okoli CO, Adams II TA, Brigljević B, Liu JJ. Design and economic analysis of a macroalgae-to-butanol process via a thermochemical route. Energy Convers Manage 2016;123:410–22.
- [177] Cechinel MAP, Mayer DA, Pozdniakova TA, Mazur LP, Boaventura RAR, de Souza AAU, et al. Removal of metal ions from a petrochemical wastewater using brown macro-algae as natural cation-exchangers. Chem Eng J 2016;286:1–15.
- [178] Henriques B, Rocha LS, Lopes CB, Figueira P, Duarte AC, Vale C, et al. A macroalgae-based biotechnology for water remediation: simultaneous removal of Cd, Pb and Hg by living *Ulva lactuca*. J Environ Manage 2017;191:275–89.
- [179] Zhang J, Hansen PK, Fang J, Wang W, Jiang Z. Assessment of the local environmental impact of intensive marine shellfish and seaweed farming—application of the MOM system in the Sungo Bay, China. Aquaculture 2009;287:304–10.
- [180] Scottish Government Report. Wild seaweed harvesting strategic environmental assessment environmental report. Scottish Government, London: APS Group Scotland; 2016.
- [181] Environment & Heritage Service. Environmentally sustainable seaweed harvesting in Northern Ireland 2007. [http://www.seaweed.ie/irish\\_seaweed/](http://www.seaweed.ie/irish_seaweed/)

- contacts/doc/seaweedharvestingniehspositionstatement.pdf (accessed April 12, 2020).
- [182] Neori A, Krom MD, Ellner SP, Boyd CE, Popper D, Rabinovitch R, et al. Seaweed biofilters as regulators of water quality in integrated fish-seaweed culture units. *Aquaculture* 1996;141:183–99.
- [183] Beveridge MCM, Phillips MJ, Macintosh DJ. Aquaculture and the environment: the supply of and demand for environmental goods and services by Asian aquaculture and the implications for sustainability. *Aquac Res* 1997;28:797–807.
- [184] Duarte CM, Holmer M, Olsen Y, Soto D, Marbà N, Guiu J, et al. Will the oceans help feed humanity? *Bioscience* 2009;59:967–76.
- [185] Butterworth A. Integrated Multi-Trophic Aquaculture systems incorporating abalone and seaweeds. Rep Nuff Aust Proj 2010.
- [186] Feng Y, Hung T, Greg K, Zhang Y, Li B, Yang J. Thermoeconomic comparison between pure and mixture working fluids of organic Rankine cycles (ORCs) for low temperature waste heat recovery. *Energy Convers Manage* 2015;106:859–72.
- [187] Brigljević B, Liu JJ, Lim H. Comprehensive feasibility assessment of a poly-generation process integrating fast pyrolysis of *S. japonica* and the Rankine cycle. *Appl Energy* 2019;254:113704.
- [188] van den Burg SWK, van Duijn AP, Bartelings H, van Krimpen MM, Poelman M. The economic feasibility of seaweed production in the North Sea. *Aquac Econ Manage* 2016;20:235–52.
- [189] Van den Burg SWK, Stuiver M, Veenstra FA, Bikker P, Contreras AML, Palstra AP, et al. A Triple P review of the feasibility of sustainable offshore seaweed production in the North Sea. Wageningen UR 2013.
- [190] Costa JC, Gonçalves PR, Nobre A, Alves MM. Biomethanation potential of macroalgae *Ulva* spp. and *Gracilaria* spp. and in co-digestion with waste activated sludge. *Bioresour Technol* 2012;114:320–6.
- [191] Tedesco S, Daniels S. Optimisation of biogas generation from brown seaweed residues: compositional and geographical parameters affecting the viability of a biorefinery concept. *Appl Energy* 2018;228:712–23.
- [192] Mushtaq F, Mat R, Ani FN. A review on microwave assisted pyrolysis of coal and biomass for fuel production. *Renew Sustain Energy Rev* 2014;39:555–74.
- [193] Zhao Y, Wang Y, Duan D, Ruan R, Fan L, Zhou Y, et al. Fast microwave-assisted ex-catalytic co-pyrolysis of bamboo and polypropylene for bio-oil production. *Bioresour Technol* 2018;249:69–75.
- [194] Borges FC, Xie Q, Min M, Muniz LAR, Farenzena M, Trierweiler JO, et al. Fast microwave-assisted pyrolysis of microalgae using microwave absorbent and HZSM-5 catalyst. *Bioresour Technol* 2014;166:518–26.
- [195] Uzoejinwa BB, He X, Wang S, Abomohra A-E-F, Hu Y, He Z, et al. Co-pyrolysis of seaweeds with waste plastics: modeling and simulation of effects of co-pyrolysis parameters on yields, and optimization studies for maximum yield of enhanced biofuels. *Energy Sources, Part A Recover Util Environ Eff* 2020;42:954–78.